

# **米SLR**

## **Evidence Evaluations for Australian Drinking Water Guidelines Chemical Fact Sheets – PFOS, PFHxS, PFOA, PFBS, and GenX Chemicals**

**PFOS, PFHxS, PFOA, PFBS, and GenX Chemicals Evaluation Report**

### **National Health and Medical Research Council**

Prepared by:

**SLR Consulting Australia** Level 11, 176 Wellington Parade, East Melbourne

VIC 3002, Australia

SLR Project No.: 640.V30693.20000

17 October 2024

Revision: 5.0

**Making Sustainability Happen** 

### **Revision Record**



### <span id="page-2-0"></span>**Basis of Report**

This report has been prepared by SLR Consulting Australia (SLR) with all reasonable skill, care and diligence, and taking account of the timescale and resources allocated to it by agreement with the National Health and Medical Research Council (the Client). Information reported herein is based on the interpretation of data collected, which has been accepted in good faith as being accurate and valid.

This report is for the exclusive use of the Client. No warranties or guarantees are expressed or should be inferred by any third parties. This report may not be relied upon by other parties without written consent from SLR.

SLR disclaims any responsibility to the Client and others in respect of any matters outside the agreed scope of the work.

### <span id="page-3-0"></span>**Executive Summary**

An Australian drinking water guideline and existing Fact Sheet are available for three perand polyfluoroalkyl substances (PFAS): for perfluorooctane sulfonic acid + perfluorohexane sulfonic acid (PFOS+PFHxS) and for perfluorooctanoic acid (PFOA). There is currently no Australian drinking water guideline or existing Fact Sheet for perfluorobutane sulfonic acid (PFBS) and hexafluoropropylene oxide ammonium salt plus hexafluoropropylene oxide dimer acid (also termed GenX Chemicals).

The National Health and Medical Research Council (NHMRC) have contracted SLR Consulting Australia Pty Ltd (SLR) to identify existing sources of guidance or guidelines on the impact of exposure to these five select PFAS in drinking water at levels higher or lower than the current Australian Drinking Water Guidelines health-based guideline values (where these exist) on human health outcomes.

An evidence scan to inform an update to the existing supporting information provided in the current Fact Sheet was also requested to be undertaken. This included levels detected in Australian drinking water, analytical/detection, monitoring and treatment guidance.

This evidence review has been undertaken in line with a new methodological framework intended to implement best practice methods for evidence evaluations as per the NHMRC Standards for Guidelines.

This Evaluation Report summarises the evaluation undertaken for the five select PFAS and concludes by identifying potential drinking water guideline values for adoption/adaption in the Australian context. The methodology of the review is also provided in more detail in an accompanying Technical Report.

The volume of information found in the literature search undertaken in August 2023 and needing to be assessed was very large. Due to resource constraints and with agreement from NHMRC with advice from the Water Quality Advisory Committee, critical evaluation of studies was prioritised to those studies that had not been previously reviewed and/or considered by an Australian agency for guidance/guideline value development. The latest review by an Australian jurisdiction in which guidance values were derived for three of the PFAS under consideration (PFOS+PFHxS and PFOA) was the document from Food Standards Australia New Zealand (FSANZ 2017b). This forms the basis of the current toxicity reference values (TRVs) for PFOS/PFHxS and PFOA which were used by NHMRC to derive the current guideline values in drinking water for these chemicals. FSANZ (2021) also published a review of immunomodulation effects, in which the jurisdiction reviewed a number of studies, findings of which are used to support discussions in this report on relevant PFAS.

The candidate drinking water guidelines (DWGs) for potential adoption/adaptation of suitable information for each of the five PFAS are provided in **Sections 6** to **10** of this report, with the conclusions presented in **Section 11**. As relevant identified guidance values have utilised different critical studies, critical effects and points of departure along with different uncertainty factors for guidance value determination, this has resulted in ranges being provided for some chemicals. In summary, the following options for guideline values were proposed.

- PFOS the current Australian health-based DWG of 70 ng/L is still considered to be appropriate.
- PFHxS a guideline value of 34 ng/L was considered as being potentially suitable (and conservative) for PFHxS on its own, as was the current Australian DWG value of 70 ng/L for the sum of PFOS + PFHxS. In practice this means it is considered



<span id="page-4-0"></span>reasonable to retain the existing guideline value of 70 ng/L as the sum of PFOS+PFHxS, with PFHxS not exceeding 34 ng/L.

- PFBS guideline values ranging from 1,041 to 2,939 ng/L in drinking water were considered as being appropriate and conservative. This would be a new DWG for this chemical.
- PFOA guideline values ranging from 9.5 to 70 ng/L in drinking water were considered as being potentially appropriate and conservative, as is the current Australian guideline value of 560 ng/L. However, due to various reasons outlined in **Sections [9.2.1](#page-72-0)** to **[9.2.5](#page-78-0)**, the confidence in the candidate guideline values (9.5 to 70 ng/L) is considered very low to low. It is therefore suggested the information is not of high enough quality to warrant revision of the current Australian guideline value for PFOA (560 ng/L), for which the confidence in the underpinning study is high.
- GenX Chemicals there is currently insufficient evidence to derive a health-based DWG for GenX Chemicals. However, a concentration of potential concern of 263 ng/L could be derived based on the limited toxicity data available. There is currently no existing DWG for GenX Chemicals.

From the available information gathered on exposure to the five PFAS of interest in Australian distributed drinking waters and the information gathered to inform supporting information in the Fact Sheet, all DWG options would be readily measurable with current commercial analytical techniques. Although existing treatment technologies do not appear to be particularly effective at removing PFAS from water, DWG options would be achievable if uncontaminated<sup>[1](#page-4-0)</sup> source waters are utilised. However, the DWG options may not be achievable for local drinking water supplies in contaminated areas without addition of a PFAS-removal treatment step or use of an alternative water supply.

<span id="page-4-1"></span>Based on concentrations identified in existing water quality data in the Australian context, it is unlikely that PFOS, PFHxS, PFBS and PFOA will present a human health risk from drinking water in uncontaminated regions of Australia. No concentrations of GenX Chemicals in drinking water were identified in the Australian context, so it is unknown if the candidate DWG proposed for GenX Chemicals will be above or below what is found in Australian drinking water. Additional research is required to identify if GenX Chemicals are found in Australian drinking water and at what levels.

[<sup>1</sup>](#page-4-1) Here uncontaminated means locations that are not directly affected by a point source of PFAS. Contaminated locations include locations where historical use of PFAS-containing firefighting foam has occurred. It is recognised that PFAS are widespread in the environment and small amounts of PFAS may still be found in uncontaminated locations.



### **Table of Contents**





### **Tables in Text**





- Table 7-1 [Potential drinking water guideline values \(ng/L\) resulting from adaptation of](#page-59-0) [PFHxS guidance values from different jurisdictions based on NTP \(2022\) critical](#page-59-0) [study as well as current Australian drinking water guideline for PFOS + PFHxS 59](#page-59-0)
- Table 8-1 [Potential drinking water guideline values \(ng/L\) resulting from adaptation of PFBS](#page-68-0) [guidance values from different jurisdictions based on two critical studies .......... 68](#page-68-0)
- Table 9-1 [Potential drinking water guideline values \(ng/L\) resulting from adaptation of PFOA](#page-82-0) [guidance values from different jurisdictions](#page-82-0) (1) .. [82](#page-82-0)
- Table 10-1 Potential drinking water guideline values (ng/L) resulting from adaptation of GenX [Chemicals guidance values from different jurisdictions based on DuPont \(2010\)89](#page-89-0)
- [Table 11-1Conclusions and DWG options from potential adoption/adaptation of suitable](#page-91-1) [information for PFOS, PFHxS, PFBS, PFOA, and GenX Chemicals .................. 91](#page-91-1)

### **Figures in Text**



### <span id="page-8-0"></span>**Acronyms and Abbreviations**







### <span id="page-11-3"></span><span id="page-11-1"></span><span id="page-11-0"></span>**1.0 Introduction and Background**

<span id="page-11-2"></span>An Australian drinking water guideline and existing Fact Sheet<sup>[2](#page-11-1)</sup> are available for three perand polyfluoroalkyl substances (PFAS): 70 ng/L for perfluorooctane sulfonic acid + perfluorohexane sulfonic acid (PFOS, CAS No. 1763-23-1 + PFHxS, CAS No. 355-46-4) and 560 ng/L for perfluorooctanoic acid (PFOA, CAS No. 335-67-1). There is currently no Australian drinking water guideline or existing Fact Sheet for perfluorobutane sulfonic acid (PFBS, CAS No. 375-73-5) and hexafluoropropylene oxide ammonium salt (CAS No 62037- 80-3) plus hexafluoropropylene oxide dimer acid (CAS No 13252-13-6) (also termed GenX Chemicals).

The National Health and Medical Research Council (NHMRC) have contracted SLR Consulting Australia Pty Ltd (SLR) to identify existing sources of guidance or guidelines on the impact of exposure to these five select PFAS in drinking water at levels higher or lower than the current health-based guideline values (where these exist) on human health outcomes.

An evidence scan to inform an update to the existing supporting information (e.g. levels detected in Australian drinking water, analysis/detection, monitoring and treatment guidance) provided in the Fact Sheet was also requested to be undertaken. The findings of this evaluation will be used by NHMRC to develop and/or update public health advice and/or health-based guideline values (if required) for inclusion in the *Australian Drinking Water Guidelines* (2011) (the Guidelines). The evidence reviews undertaken by SLR were governed by a newly designed methodological framework intended to implement best practice methods for evidence evaluations as per the *2016 NHMRC Standards for Guidelines*. For each PFAS, SLR was asked to:

- Customise and apply the 'Research Protocol' template provided by NHMRC to answer research questions.
- Produce a Technical Report and an Evaluation Report for five select PFAS.
	- o The Technical Report is to capture the details and methods used to undertake each review.
	- o The Evaluation Report is to interpret, synthesise and summarise the existing guidance and evidence pertaining to the research questions.

These tasks were performed in consultation with NHMRC's Water Quality Advisory Committee (the Committee) and NHMRC.

For the five select PFAS, the requirements of the evaluation were as follows:

- <span id="page-11-4"></span>1 Screen any existing guidance/guidelines<sup>[3](#page-11-3)</sup> (if available).
- 2 Collate and review any useful supporting information for modification/expansion of the existing PFAS (PFOS+PFHxS and PFOA) chemical Fact Sheet.

 $3$  A guidance value is the same as a Toxicity Reference Value (TRV) and refers to a health-based intake of a chemical which can be ingested daily over a lifetime without adverse health effects. A guideline value for various environmental media (including drinking water) uses the health-based guidance value in its derivation but may only apportion a certain percentage of the guidance value to the intake from that particular medium.



<sup>&</sup>lt;sup>[2](#page-11-2)</sup> A single Fact Sheet currently exists for PFOS+PFHxS and PFOA (NHMRC and NRMMC 2011); advice on new chemicals would either be included in the same Fact Sheet or new Fact Sheets developed as required if determined by NHMRC with advice from the Committee.

The report herein is the Evaluation Report for the five PFAS evaluated (PFOS, PFOA, PFHxS, PFBS and GenX Chemicals). A combined Evaluation Report was produced since there was a large cross-over between the information for the various PFAS evaluated.

### <span id="page-12-0"></span>**1.1 Objectives**

The overarching objective of this review is to identify relevant information from existing guidance/guidelines on the impact of exposure to each of the five select PFAS (i.e. PFOS, PFHxS, PFOA, PFBS, and GenX Chemicals) in drinking water on human health outcomes.

Another objective of the review is to undertake an evidence scan to inform any modification/expansion of supporting information (e.g. monitoring and treatment guidance) that is provided in the existing PFAS Fact Sheet.

### <span id="page-12-1"></span>**2.0 Research Questions**

Research questions for this review were drafted by SLR and peer reviewed and agreed upon by the Committee and NHMRC prior to conducting the literature searches. The research questions guiding the review are provided in **[Table 2-1](#page-12-2)**.

<span id="page-12-2"></span>





### <span id="page-13-0"></span>**3.0 Methodology Overview**

As part of the review, a number of literature searches were undertaken to target specific information relevant to answering the research questions. They consisted of the following:

 A targeted literature search undertaken in August 2023 of existing health-based guidance/guidelines. Jurisdictions included in this search were those previously identified by ToxConsult (2019) as providing reliable information and meeting a large proportion of pre-determined technical and administrative criteria as per the Assessment Tool in the Technical Report. They included the World Health Organization (WHO) including the Joint FAO/WHO Expert Committee on Food Additives (JECFA), European Food Safety Authority (EFSA), United States Environmental Protection Agency (US EPA), US Agency for Toxic Substances and Disease Registry (ATSDR), Californian Office of Health and Hazard Assessment (OEHHA), Food Standards Australia New Zealand (FSANZ), and the Australian Pesticides and Veterinary Medicine Authority (APVMA).



- <span id="page-14-0"></span> As it was known prior to undertaking the search that other jurisdictions (not identified in the first dot point above) had also recently derived guidance/guideline values for the five PFAS under consideration, a number of additional jurisdictions were included in the search. These were Health Canada, Dutch National Institute for Public Health and the Environment (RIVM), German Bundesinstitut für Risikobewertung (BfR – Federal Institute for Risk Assessment), US Centre for Disease Control (CDC), Australian Industrial Chemicals Introduction Scheme (AICIS), and various US state health departments including Minnesota, Washington, Maine, Alabama, Alaska, Connecticut, Vermont, New Jersey, Michigan and Massachusetts.
- An additional evidence scan of recent publicly available literature for supporting information in the Fact Sheet (e.g. general description, uses, measurement techniques and limits of reporting in drinking water, treatment options, etc.).

Results were subjected to the following steps in order to identify the most relevant information:

- A preliminary title screen where titles of results were scanned by a researcher and a decision recorded regarding relevance of the result; and
- A content screen where full text content of reports/reviews/articles selected to be included from the preliminary title screen step were reviewed in relation to the research questions by a subject expert to determine which to include in data extraction.

Relevant data were extracted by populating various pre-constructed tables which focused on data needed to answer the research questions. Synthesis was conducted by presenting summarised extracted data in tabular format for each individual research question. For each candidate jurisdiction's guidance/guideline value identified for the five PFAS included in this report, an evaluation of existing jurisdiction Guidelines was undertaken with respect to a defined list of administrative and technical criteria (previously defined by ToxConsult 2019 and NHMRC) using an Assessment Tool. The reader is referred to the accompanying Technical Report for the detailed methodology, records of the literature screening process (including all records that were excluded) and all data extraction, and Assessment Tool tables.

<span id="page-14-1"></span>The volume of information found and needing to be assessed was very large. Due to resource constraints and with agreement from NHMRC with advice from the Committee, critical evaluation of studies underpinning existing guideline values in this Evaluation Report was prioritised to those studies that had not been previously reviewed and/or considered by an Australian agency for guidance/guideline value development. The latest review by an Australian jurisdiction in which guidance values were derived for three of the PFAS under consideration (PFOS+PFHxS and PFOA) was the Food Standards Australia New Zealand (FSANZ 2017b)<sup>[4](#page-14-0)</sup> document. This forms the basis of the current toxicity reference values (TRVs) for PFOS/PFHxS and PFOA which have been used by NHMRC to derive the current guideline values in drinking water for these chemicals. FSANZ (2021) also published a review of immunomodulation effects, in which the jurisdiction reviewed a number of studies, findings of which are used to support discussions in this report on relevant PFAS. This agreed amendment to the scope of the Evaluation Report is captured in an addendum to the Research Protocol (see **Section 3.4** in Technical Report). This Evaluation Report provides the following.

[<sup>4</sup>](#page-14-1) Based on an evaluation of the 'must-have', 'should-have' and 'may-have' administrative and technical criteria specified in the Assessment Tool in the Research Protocol, it is concluded that the FSANZ (2017b) quidance would be suitable for adoption/adaptation (see details in Technical Report).



- <span id="page-15-0"></span> A tabular summary of the various guidance/guideline values found in the literature review (and for which data extraction summaries are provided in the Technical Report). This tabular summary in **Section [5.0](#page-41-0)** provides colour coding for the health endpoints on which the guidance/guideline values are based.
- The full list of critical studies underpinning each of the quidance values derived by various national and international jurisdictions is shown in **Appendix A** of this Evaluation Report, along with an indication of whether or not the critical study had been previously evaluated / considered by FSANZ (2017b, 2021).
- <span id="page-15-1"></span> Discussions/critical evaluation of those studies underpinning existing guidance values not previously considered in the FSANZ (2017b) review. These are the studies marked with a cross (i.e. 'x') in **Appendix A** in the column denoted 'FSANZ (2017b)'. In line with the agreed change to the scope of the Evaluation Report, critical review of guidance values in this Evaluation Report was limited to the following.
	- o All the GenX Chemicals and PFBS guidance values (as these two PFAS were not previously considered by an Australian agency).
	- o For PFOS:
		- Values derived by US EPA (critical study: Budtz-Jørgensen and Grandjean 2018).
		- Value for PFOS, PFOA, PFHxS derived by EFSA (critical study: Abraham et al. 2020).[5](#page-15-0)
	- o For PFOA:
		- Value derived by OEHHA (critical studies: Gallo et al. 2012; Li et al. 2017).
		- **Value derived by ATSDR (critical study: Koskela et al. 2016).**
		- Value derived by New Jersey Department of Environmental Protection (NJDEP) (critical study: Loveless et al. 2006).
		- Value derived by Michigan PFAS Action Response Team (MPART) (critical studies: Onishchenko et al. 2011, Koskela et al. 2016).
		- Value for PFOS, PFOA, PFHxS derived by EFSA (critical study: Abraham et al. 2020).
		- Value for PFOA derived by US EPA (critical study: Budtz-Jørgensen and Grandjean 2018).
	- o For PFHxS:
		- Value for PFOS, PFOA, PFHxS derived by EFSA (critical study: Abraham et al. 2020).
		- Value derived by Minnesota Department of Health (MDH), MPART and OEHHA (critical study: NTP 2022).
- A brief summary of supporting information was provided in the Evaluation Report, with further detail provided in the Technical Report if required by NHMRC and the Committee.

**Figure 1** shows an overview of the literature search process followed for the five PFAS included in this review. This is presented as a PRISMA (Preferred Reporting Items for Systematic Reviews and Meta-Analyses) flow diagram that describes the study selection process and numbers of records at each stage of screening (Moher et al. 2009).

<sup>&</sup>lt;sup>[5](#page-15-1)</sup> Note this study has since been evaluated by FSANZ (2021); the FSANZ (2021) evaluation of the study was primarily relied upon along with information from other jurisdictions that have considered use of this information in derivation of their guidance/guideline values.



<span id="page-16-0"></span>



\*Some reviews derived guidance/guideline values for more than one PFAS.

# This value indicates the number of agency reviews that data was extracted from for each individual PFAS as shown in Appendix B of the Technical Report. Not all agency reviews had guideline values/guidance as some were used for supporting information only. Due to resource constraints and with agreement from NHMRC with advice from the Committee, critical evaluation of studies underpinning existing guideline values in this Evaluation Report was prioritised to those studies that had not been previously reviewed and/or considered by an Australian agency for guidance/guideline value development (see **Appendix A**).



This report provides the summary of the findings (**Section [4.0](#page-17-0)**), a tabular summary of all existing guidance/guideline values sourced in the literature search (**Section [5.0](#page-41-0)**), a discussion of the results for each PFAS (**Section [6.0](#page-48-0)** to **[10.0](#page-86-0)**), and a conclusion (**Section [11.0](#page-91-0)**). Where health-based information was considered reasonable for potential derivation of a guideline value, calculations of prospective drinking water guidelines (DWGs) were undertaken using the methodology and default assumptions outlined in the Guidelines (NHMRC and NRMMC 2011) unless otherwise advised by the Committee.

The default equation is outlined in NHMRC and NRMMC (2011, Section 6.3.3) and has been adapted below as Equation 1. In this instance, units have been added in to show how they cancel out and the 'animal dose' in the equation can in fact be an animal or human dose, since both data types may be used to derive DWGs. In some instances, if adaptation of existing guidance values was considered, these guidance values may already incorporate the safety factor shown in the denominator of Equation 1.

```
Guideline value (ng/L) =
```

```
animal or human dose (ng/kg bw/d) x human weight (kg bw) x proportion of intake from water (fraction)
          volume of water consumed (L/d) x safety factor (unitless)
```
Default assumptions typically used in the Guidelines are 70 kg bw for adult human body weight (or 13 kg bw for 2-year old child or 5 kg for an infant), 10% (0.1) for the proportion of intake from drinking water (apart from bottle-fed infants, where 100% is used), and 2 L/day of water consumption by an adult (1 L/day by a child, 0.75 L/day by a bottle-fed infant).

### <span id="page-17-0"></span>**4.0 Results**

The targeted screening of existing health-based guidance identified multiple existing healthbased guidance/guideline values for the five PFAS included in this evaluation in the literature consulted. Responses to research questions were informed by the data extractions from the guidance/guideline documents found in the literature reviewed.

Detailed summary findings tables for each research question are provided in the Technical Report. In this Evaluation Report, the research question tables have been condensed to highlight differences between the various agency guidance/guideline documentation and other studies where they have been identified.

### <span id="page-17-1"></span>**4.1 Health-based aspects**

Research questions 1-8 all cover health-based aspects of the review; this is considered to be the most important information in the Fact Sheet. **[Table 4-1](#page-18-0)** provides a synthesis of the results.

### **Table 4-1 Summary of findings from data extraction for health-based aspects**

<span id="page-18-0"></span>




















































# **4.2 Exposure-related aspects**

Another important aspect of the Fact Sheet covers exposure-related considerations. This is important for consideration of whether exposures by Australians to the chemicals evaluated are potentially approaching a health-based guidance value that will be used for deriving a candidate DWG. It is also important for considerations of whether typical levels of the chemical considered in Australian drinking water supplies would adhere to any derived DWG. Research questions 9-11 cover exposure-related aspects of the review. For these aspects, drinking water quality reports from various water corporations around Australia were consulted in addition to the jurisdictional reviews sourced as part of the health-based review and the supporting information. **[Table 4-2](#page-36-0)** provides a synthesis of the results.

<span id="page-36-0"></span>





## **4.3 Risk-based aspects**

Research questions 12 and 13 are risk-based considerations. The publications subjected to detailed data extraction were also consulted to answer these questions. **[Table 4-3](#page-37-0)** presents a summary of the findings.

<span id="page-37-0"></span>**Table 4-3 Summary of findings from data extraction for risk-based research questions**

#	Research Questions	<b>Findings</b>
	What are the risks to human health from	Provided drinking water catchments are protected from PFAS contamination and alternative water supplies are available if



# **4.4 Supporting information**

Supporting information in Fact Sheets for chemicals in the Guidelines typically consist of (NHMRC and NRMMC 2011) a brief general description (i.e. uses of PFAS, sources in drinking water), typical values in Australian drinking water, treatment of drinking water, and measurement (i.e. analytical) considerations. The remaining Research questions 14-24 cover the supporting information of the review. For these aspects, in addition to consulting the previously mentioned sources (e.g. the drinking water quality reports from various water corporations and utilities around Australia, the health-based jurisdictional literature identified in the targeted search), additional targeted searches were undertaken (for details, refer to Technical Report). **[Table 4-4](#page-39-0)** provides a summary of the results.

### <span id="page-39-0"></span>**Table 4-4 Summary of findings from data extraction for supporting information**







spectrometry. TOP = Total Oxidisable Precursor Assay. TOF Assay = Total Organic Fluorine Assay.

# <span id="page-41-0"></span>**5.0 Tabular summary of existing guidance/guideline values**

It is noted guidance/guideline values differ quite markedly depending on the jurisdiction and there is discrepancy with respect to what jurisdictions consider to be the critical health endpoint. Guidance/guideline values have also been reducing rapidly over time as new studies and new interpretation of the data emerge in the publicly available literature. To gain a visual appreciation of the differences and the lowering of values over time, **[Table 5-1](#page-43-0)** and **[Table 5-2](#page-46-0)** provide tabular summaries of the guidance and guideline values, respectively, derived by the various jurisdictions for PFOS, PFHxS, PFBS, PFOA and GenX Chemicals. It is noted only the most recently derived guidance/guideline values were subjected to data extraction in the Technical Report. Older values are nevertheless provided in the tables and



were sourced from the various jurisdictional websites for historical context. The summary tables are colour coded according to the critical health endpoint that underpins the guidance/guideline value.

From **[Table 5-1](#page-43-0)**, it is evident there is much disparity between the critical health endpoints chosen by the various jurisdictions for derivation of guidance values for PFOS and PFOA, and less so for PFHxS, whereas for PFBS and GenX Chemicals most jurisdictions that have derived a guidance value seem to concur with respect to the critical health endpoint (although the latter is possibly due to the relative lack of available studies for deriving guidance/guideline values). There is even more variability between resulting guideline values, as shown in **[Table 5-2](#page-46-0)**, due to some not being health-based and there being wide variation in the assumptions used to derive them. Due to the large variability in guideline values, it was deemed most informative to focus on critically reviewing available guidance values.

<span id="page-43-0"></span>

#### **Table 5-1 Summary of existing guidance values for the five PFAS included in this report (ng/kg/day)**

JП





- (a) A TRV was not derived by WHO (2022a). A DWG was derived based on pragmatism.
- (b) Health-based guidance value from EFSA (2020a) applies to the sum of four PFAS, i.e. ∑PFOA, PFNA, PFHxS and PFOS.
- $(c)$  A TRV for PFOS was conservatively adopted for PFHxS. The TRV is applied as a sum (PFOS+PFHxS).
- (d) The TRV from ATSDR is based on intermediate exposure timeframe (14 <365 days).
- (e) Based on a relative potency factor of 0.06 for GenX, 0.001 for PFBS and a TRV of 12.5 ng/kg/day (for PFOA). From 2021, the TRV for PFOA (summed with three other PFAS) may be substantially lower (0.63 ng/kg/day).
- (f) It is presumed by SLR that BfR adopted the updated tolerable weekly intake in 2020 from EFSA (2020a) given that BfR adopted the 2018 value from EFSA (2018, as quoted in BfR 2019a).
- (g) These guidance values are based on studies not previously evaluated/considered by FSANZ (2017b) and have been further evaluated in **Sections [6.0](#page-48-1)** to **[10.0](#page-86-1)**.

#### **Legend:**



### <span id="page-46-0"></span>**Table 5-2 Summary of existing drinking water guideline values for the five PFAS included in this report (ng/L)**





17 October 2024 SLR Project No.: 640.V30693.20000



(c) Reference Levels (RLs) and health protective concentration (HPC) for non-cancer effects shown. The RLs and Public Health Goal (PHG) for cancer effects are not shown. In 2019, State Water Resources Control Board (SWRCB) (NLs) at the lowest levels at which PFOA and PFOS can be reliably detected in drinking water (OEHHA 2019a).

(e) "Any change to a [State Action Level] SAL or adopting a state [Maximum Contaminant Level] MCL requires rulemaking. WSDH will continue to implement SALs until rulemaking permits use of this value (a MCL from USEPA)" (f) Health-based water concentration (HBWC) are the "acceptable" values used to create a ratio of observed/acceptable for each of 4 PFAS (PFNA, PFHxS, PFBS and GenX). If the ratios add up to more than 1.0, action must be t drinking water.

Department of Public Health/Department of Environmental Protection. CDPH = Connecticut Department of Public Health. US EPA = United States Environmental Protection Agency.

(a) A DWG was derived based on pragmatism. A value of 500ng/L is applicable to Total PFAS.

(b) The DWG is applied as a sum: PFOS+PFHxS.

(d) Drinking water guideline shown is for a child (which is lower than the value derived for an adult).

(g) Interim State drinking water standard for the combined sum of six different PFAS: PFOA, PFOS, PFHpA, PFNA, PFDA, and PFHxS.

(h) A 0.07 µg/L action level was set for the sum of the following five (5) PFAS chemicals: PFOS, PFOA, PFNA, PFHxS, and PFHpA.

(i) In order to align state actions to the recently announced EPA plans, Alaska DEC will use the US EPA Lifetime Public Health Advisory (LHA) (PFOS+PFOA above 0.07 µg/L) as the Action Level.





# <span id="page-48-1"></span><span id="page-48-0"></span>**6.0 Discussion for PFOS**

This section provides a discussion of the strengths and limitations of the identified guidance values for PFOS for possible adoption/adaptation into the Guidelines. Critical evaluation was focused on those guidance values derived using underpinning studies not previously considered / evaluated by FSANZ (2017b). FSANZ (2021) also published a review of immunomodulation effects, in which the jurisdiction reviewed a number of studies, findings of which have been used to support subsequent discussions.

## <span id="page-48-2"></span>**6.1 Potential suitability of health-based guidance values for possible adoption/adaptation**

Candidate guidance values for PFOS described in **Section [4.1](#page-17-0)** for possible adoption/adaptation in Australia have been evaluated using the Assessment Tool provided in Appendix D in the Technical Report. This tool evaluates each document against administrative and technical criteria that demonstrate transparent and robust guideline development and evidence review processes that meet NHMRC standards for guidelines. The overall potential suitability of the guidance values for adoption/adaption can be gauged at least partially by examining the percentage of 'must-have', 'should-have', and 'may-have' criteria met by each jurisdiction.

**[Figure 6-1](#page-49-1)** presents the percentage of criteria (combined technical and administrative criteria) met by each jurisdiction. It is evident from the figure that several publications met similar percentages of criteria, with ATSDR (2021a), EFSA (2020a), FSANZ (2017b), NJDEP (2019b), OEHHA (2023a), and US EPA (2022c, e; 2021b) all meeting relatively high (i.e. ~>60%) proportions of 'must-have' and 'should-have' criteria.

Other jurisdictions (HC 2018a, MDH 2020a, MPART 2019a, OEHHA 2019a) met lower proportions of criteria, indicating these guidance documents potentially do not conform with modern methods of undertaking systematic reviews.



- <span id="page-49-1"></span>**Figure 6-1 Overall proportion of 'must-have', 'should-have' and 'may-have' technical/administrative criteria as per the Assessment Tool met by jurisdictions who have derived candidate guidance values for PFOS for possible adoption/adaptation in Australia**
- <span id="page-49-0"></span>**6.2 Critical evaluation of PFOS guidance values based on underpinning studies not previously considered by FSANZ (2017b)**

For PFOS, the only guidance values identified in the literature review that based their derivations on underpinning studies not previously considered / cited in the comprehensive review undertaken by FSANZ (2017b) are the following. The discussion in this section therefore focuses on the relevant studies underpinning these guidance values.

- EFSA (2020a) who derived a guidance value for ∑PFOA, PFNA, PFHxS and PFOS of 0.63 ng/kg bw/day (TWI = 4.4 ng/kg bw per week). The critical study this was based on is Abraham et al. (2020), which is a cross-sectional study of 101 healthy 1-year old children which found statistically significant inverse associations between serum levels of PFOA, but not of PFOS, and adjusted levels of vaccine antibodies against *Haemophilus influenza* type b (r = -0.32), diphtheria (r = -0.23) and tetanus ( $IqG1$  only) ( $r = -0.25$ ). When subjects were stratified according to PFOA concentration, comparison of the highest and lowest quintiles showed that PFOA was associated with antibody levels, on a logarithmic scale, that were 86% lower for *Haemophilus influenza* type b, 53% lower for diphtheria and 54% lower for tetanus. This effect is a marker of immune response. The EFSA CONTAM Panel decided to combine its assessment on the sum of four PFAS, i.e. PFOA, PFNA, PFHxS and PFOS as these four PFAS contribute most to the levels observed in human serum, share toxicokinetic properties in humans and show similar accumulation and long half-lives (EFSA 2020a).
- US EPA (2022c, e; 2021b) who derived a DRAFT guidance value of 0.0079 ng/kg/day for PFOS based on decreased antibody titre following diphtheria vaccination in 1-year old children – also a marker of immune response - in studies



<span id="page-50-0"></span>by Grandjean et al. (2012) and Budtz-Jørgensen and Grandjean (2018). It is noted the former study (Grandjean et al. 2012) was previously considered by FSANZ (2017b) but was not selected for derivation of a guidance value.

### **6.2.1 Abraham et al. (2020) – used by EFSA (2020a)**

FSANZ (2021) recently undertook an updated review of PFAS and immunomodulation, in which they provided a critique of the Abraham et al. (2020) study. Strengths of the study, according to FSANZ (2021) included the following.

- *"Children are very close in age.*
- *The investigations of immune parameters were relatively thorough.*
- *Some other persistent organic pollutants were considered.*
- *Differences between breastfed and formula-fed children were considered.*
- *Because the samples were collected in the 1990s, higher PFAS levels were present than in more recent studies."*

Limitations of the study, according to FSANZ (2021), included the following.

- "*The cohort size was very small, only 101 children overall.*
- *There is substantial interindividual variability in response.*
- *There is a lack of information on whether the decreases in antibody concentrations are clinically relevant. That is, PFOA may cause antibody titres to fall below effective levels sooner than they naturally would have, but if the recommended vaccine schedule is followed, antibody titres might remain sufficient to protect against disease, particularly in formula-fed infants.*
- *The question of the stability of antibodies in samples stored for decades is not addressed."*

EFSA (2020a) themselves noted similar limitations associated with use of the endpoint identified in the Abraham et al. (2020) study for guidance value development. Other submitters' comments on the draft EFSA (2020a) document, according to FSANZ (2021) included the following.

- "*The associations in the studies considered pivotal by the EFSA CONTAM Panel are weak, and cross-sectional studies cannot demonstrate causation.*
- *Vaccination response in humans is known to be highly variable, and the decline in antibodies after vaccination is not well defined but cannot be assumed to be linear.*
- *The mechanism/s by which PFAS affect the immune system are poorly understood.*
- *It is not appropriate to apply Physiologically Based Pharmacokinetic (PBPK) models validated for adults to data obtained from breastfed infants or small children.*
- *It is not appropriate to derive a TRV for adults from data from breastfed infants.*
- *Other authoritative bodies have identified different critical effects for the individual PFAS.*
- *It is clear from the available data that the potencies of the four PFAS included in the guidance value from EFSA (2020a) differ*."

In addition to the limitations identified by FSANZ (2021) and the submitters' comments on the draft EFSA (2020a) document, it becomes clear from the scatter plots of the data for combined PFAS as shown in Appendix K of the EFSA (2020a) report and reproduced in **[Figure 6-2](#page-51-0)** below that there is wide spread in the data and any suggestive inverse association (as shown on the graphs with the red lines) appears to be markedly influenced by the few data points in the 50-60 µg/L serum PFAS range. Thus, the association may partially be an artefact of not having enough data in the highest quintile.



Note: Broad grey band = moving average; red line = fitted 'knee' function; horizontal green line = mean minus one standard deviation of the antibody levels below the 'knee'; vertical grey line = PFAS sum level of the 'knee'; vertical blue line = PFAS sum level of the 'knee' function with antibody levels averagely diminished by one standard deviation.

#### <span id="page-51-0"></span>**Figure 6-2 Scatter plot of levels of vaccine antibodies (K.1** *Haemophilus influenza* **type b, K.2 Tetanus, K.3 Diphtheria) in relation to the sum of PFAS (PFOA, PFNA, PFHxS, and PFOS) in serum (reproduced from Appendix K in EFSA 2020a)**

In the toxicological profile for PFAS from ATSDR (2021a), the agency remarks that there are sufficient epidemiological data to identify possible sensitive targets for many of the PFAS; however, there are two major limitations to establishing dose-response relationships for these effects and using the epidemiological studies to derive TRVs: i) accurate identification of environmental exposure levels producing increased risk for adverse effects (exposure estimates and routes of exposure) and ii) likely co-exposure to mixtures of PFAS. Other limitations include the cross-sectional design of the majority of epidemiological studies and the potential that reverse causality contributes to the observed associations. Although the epidemiological databases for several PFAS provide valuable information on hazard identification, ATSDR (2021a) considered the uncertainties regarding doses associated with adverse effects and possible interactions between compounds preclude use of these data to derive TRVs.

FSANZ (2021) concluded that consistent with earlier observations, the associations between PFAS blood levels and antibody titres in the Abraham et al. (2020) study as well as other studies included in the review were generally weak and partially inconsistent findings have been observed for PFOS, PFOA and other PFAS for the same antigen. It was concluded by FSANZ (2021) while these studies provide limited evidence of statistical associations, "*a causal relationship between increased PFAS blood levels and impaired vaccine response cannot be established with reasonable confidence. The evidence for an association between increasing PFAS blood levels and impaired vaccine response is insufficient for quantitative risk assessment for a number of reasons which include the following*.

- *The small number of studies and participants, and mostly cross-sectional design of studies such that conclusions around causality should be drawn with caution.*
- *Limited dose-response information with most studies investigating a narrow range of blood levels associated with background levels of PFAS exposure.*
- *Inconsistency in antibody response to vaccines between different PFAS congeners which cannot explained by study design.*
- *Potential for confounding by other known environmental immunotoxicants such as PCBs for which inverse associations with blood serum antibody concentrations against tetanus and diphtheria have previously been reported living in the Faroe islands.*



<span id="page-52-0"></span> *Uncertainty about the clinical relevance, if any, of the observed statistical associations to susceptibility to infectious disease."*

This lines up with the information in the Australian Immunisation Handbook (DHAC 2018) that vaccine effectiveness can be assessed in a number of ways including by assessing the following.

- "*How effective the vaccine is at preventing infection.*
- *How effective the vaccine is at preventing hospitalisation for the disease.*
- *The impact of a vaccination program on disease incidence in the population*."

A reduction in antibody titre response, whilst a potential marker of immune response, does not appear to be readily correlated with an adverse response *per se.*

HC (2018a) also commented on the clinical importance in humans of the endpoint under discussion. They cited a study by Grandjean et al. (2012), which, according to HC (2018a) demonstrated that despite decreased vaccine-specific immunoglobulin response in PFOSexposed children, the number of children with immunoglobulin levels below the clinically protective level was low. They also stated that in humans, evidence of immunosuppression is inconsistent, and the influence of PFOS exposure on clinical immunosuppression (i.e. incidence of illnesses) appears to be more tenuous. Therefore, although low PFOS doses appear to be associated with immunosuppression, the data are not considered to be reliable for use as a key study for derivation of a TRV.

#### <span id="page-52-2"></span>**6.2.2 Budtz-Jørgensen and Grandjean (2018) – used by US EPA (2022c, e; 2021b)**

The authors of the Budtz-Jørgensen and Grandjean (2018) study undertook a benchmark dose analysis on a prospective birth Faroe Islands cohort from previous studies by the same research group (Grandjean et al. 2012, 2017) on associations of serum PFAS with vaccine antibody concentrations. Grandjean et al. (2012) was previously considered / reviewed by FSANZ (2017b) in derivation of the TRVs for PFOS and PFOA. The Grandjean et al. (2017) study was also included in the FSANZ (2021) review when the agency cited a systematic literature review by Kirk et al. (2018) which was conducted as part of the PFAS Health Study in Australia.<sup>[6](#page-52-0)</sup> The main study findings from the Kirk et al. (2018) systematic literature review with respect to immunomodulatory effects of PFAS were as follows.

<span id="page-52-1"></span> For diphtheria vaccine, there was limited evidence for an association between PFOA, PFOS, PFHxS and PFDA, noting that three of the four papers reviewed by Kirk et al. (2018) were on the same cohort in the Faroe Islands.

<sup>&</sup>lt;sup>[6](#page-52-1)</sup> The PFAS Health Study was commissioned by the Australian Government and was undertaken by the National Centre for Epidemiology and Population Health at the Australian National University (ANU) (https://nceph.anu.edu.au/research/projects/pfas-health-study). The study investigated the exposure levels and potential health effects of PFAS in areas of known contamination in the communities of Williamtown in New South Wales, Oakey in Queensland, and Katherine in the Northern Territory, Australia. Areas in these places have been contaminated with PFAS due to firefighting activities on nearby Defence Force bases. The study found that blood levels of PFOS and PFHxS were elevated in the exposed communities compared to comparison communities. It also found that there were higher levels of psychological distress among people in exposed communities. Higher PFAS levels in blood were associated with higher blood cholesterol in people from Williamtown, and higher uric acid levels in people from Williamtown and Katherine. The effects are small and unlikely to lead to poor health outcomes (ANU 2022). The study found no association with the other health outcomes investigated. The PFAS Health study also found that main risk factors for elevated blood concentrations of PFAS were the length of residence in an exposed community, at least weekly consumption of bore water or certain locally grown foods, and occupational exposure to firefighting foams.



- For response to rubella vaccine, the evidence for an association was limited for PFOA and PFOS, and inadequate for PFHxS and PFNA.
- For all other vaccines (tetanus, measles, mumps and influenza), the evidence for an association was inadequate.
- With regard to associations between PFAS exposure and adverse health outcomes, the evidence for all health outcomes considered (hospitalisations due to infection, middle ear infection, gastroenteritis and colds/influenza) was inadequate.
- The evidence for adverse effects of PFAS on all allergy and hypersensitivity endpoints, including asthma, allergies (including food allergies), plant sensitivity, shrimp allergy, cockroach sensitivity, mould sensitivity, allergic rhinoconjunctivitis, wheezing and eczema, was inadequate.

Many of the same comments made in **Section [6.2.1](#page-50-0)** also apply to the use of this study for guidance value derivation.

## <span id="page-53-1"></span>**6.3 Summary and Conclusion for PFOS**

Although ten health-based guidance values for potential adoption/adaptation were sourced from international jurisdictions reviewed for this report, only two of these used data in the derivation that had previously not been considered / evaluated by FSANZ (2017b).

These were the EFSA (2020a) and US EPA (2022c, e; 2021b) guidance values for PFOS, which used two studies to underpin the derivation that had not been previously considered / evaluated by FSANZ (2017b), i.e. Abraham et al. (2020) and Budtz-Jørgensen and Grandjean (2018).

Based on a brief critical evaluation of the two studies, consistent with the conclusions made by FSANZ (2021), it is concluded that a causal relationship between increased PFAS serum levels and impaired vaccine response cannot be established with reasonable confidence from the available human epidemiological information. The evidence for an association between increasing PFAS serum levels and impaired vaccine response is insufficient for the endpoint to be used for derivation of PFOS TRVs.

It is therefore concluded the current Australian guidance value for PFOS of 20 ng/kg/day and DWG value for PFOS + PFHxS of 70 ng/L are still appropriate. The derivation of these values is briefly shown in **[Table 6-1](#page-53-0)** below.



#### <span id="page-53-0"></span>**Table 6-1 Derivation of current Australian drinking water guideline value (ng/L) for PFOS (NHMRC and NRMMC 2011; FSANZ 2017b; DOH 2017)**

<span id="page-54-0"></span>

of a short-term study; UF<sub>composite</sub> = Composite (i.e. total) uncertainty factor; UF<sub>database</sub> = Uncertainty factor to account for the limited database of toxicological studies.

(1) NHMRC and NRMMC (2011) followed the default assumptions for derivation of DWGs in Australia using the following equation:

<span id="page-54-1"></span>DWG (ng/L) = [Guidance value (ng/kg bw/day) x 70kg (adult) x 0.1 for adult]  $\div$  2 L/day for adult

Concentrations of PFOS in uncontaminated distributed drinking water in Australia can range up to 6 ng/L in Queensland (QAEHS 2018a, 2018b)<sup>[7](#page-54-0)</sup> and Sydney (Sydney Water 2023) but up to 16 ng/L in Australia according to WHO (2022). PFOS+PFHxS concentration was found to be at 90% of the Australian DWG (i.e. ~60 ng/L) in one bore in a drinking water borefield supplying Esperance, Western Australia (WCWA 2019, 2020). Once this apparent PFOS/PFHxS contamination was identified, this bore was no longer used (WCWA 2023). Thus, PFOS is unlikely to present a human health risk from uncontaminated distributed drinking water in most regions of Australia. However, there are many sites of PFAS contamination in Australia, and, if water from these contaminated sites is used as a local source of drinking water (e.g. backyard bore in rural location where distributed water is not available), PFOS may be present at concentrations greater than the existing Australian DWG (and therefore also the candidate DWG suggested in this report) in these cases.

# **7.0 Discussion for PFHxS**

This section provides a discussion of the strengths and limitations of the identified guidance values for PFHxS for possible adoption/adaptation into the Guidelines. Critical evaluation

<sup>&</sup>lt;sup>[7](#page-54-1)</sup> Note the Queensland data is for raw water catchments.

was focused on those guidance values derived using underpinning studies not previously considered / evaluated by FSANZ (2017b).

### <span id="page-55-1"></span>**7.1 Potential suitability of health-based guidance values for possible adoption/adaptation**

Candidate guidance values for PFHxS described in **Section [4.1](#page-17-0)** for possible adoption/adaptation in Australia have also been evaluated using the Assessment Tool provided in Appendix D in the Technical Report and already described in **Section [6.1](#page-48-2)** for PFOS.

**[Figure 7-1](#page-55-0)** presents the percentage of criteria (combined technical and administrative criteria) met by each jurisdiction. It is evident from the figure that several publications met similar percentages of criteria, with ATSDR (2021a), EFSA (2020a), FSANZ (2017b), OEHHA (2022a), and US EPA (2023) all meeting relatively high (i.e. ~>60%) proportions of 'must-have' and 'should-have' criteria.

Other jurisdictions (MDH 2020b, MPART 2019a) met lower proportions of criteria, indicating these guidance documents potentially do not conform with modern methods of undertaking systematic reviews.



- <span id="page-55-0"></span>**Figure 7-1 Overall proportion of 'must-have', 'should-have' and 'may-have' technical/administrative criteria as per the Assessment Tool met by jurisdictions who have derived candidate guidance values for PFHxS for possible adoption/adaptation in Australia**
- **7.2 Critical evaluation of PFHxS guidance values based on underpinning studies not previously considered by FSANZ (2017b)**

For PFHxS, the only guidance values identified in the literature review that based their derivations on underpinning studies not previously considered / cited in the comprehensive



<span id="page-56-0"></span>review undertaken by FSANZ (2017b) are the following. The discussion in this section therefore focuses on the relevant studies underpinning these guidance values.

- EFSA (2020a) who derived a guidance value for ∑PFOA, PFNA, PFHxS and PFOS of 0.63 ng/kg bw/day (TWI = 4.4 ng/kg bw per week) for decreased antibody titre to specific vaccines in children. The critical study underpinning this guidance value has already been critically evaluated in **Section [6.2.1](#page-50-0)**.
- US EPA (2023) who derived a DRAFT guidance value of 0.0004 ng/kg/day for PFHxS based on decreased antibody titre following diphtheria vaccination in 1-year old children. The critical studies underpinning this guidance value have already been critically evaluated either by FSANZ (2017b) (in the case of Grandjean et al. 2012) or in **Section [6.2.2](#page-52-2)** (in the case of Budtz-Jørgensen and Grandjean 2018).
- <span id="page-56-1"></span> Three US State jurisdictions (MDH 2020b, MPART 2019a, OEHHA 2022a) all derived a TRV of 9.7 ng/kg/day for PFHxS based on decreased thyroxine (T4) in rats. The critical study underpinning this derivation is NTP (2022)<sup>[8](#page-56-0)</sup> which was not previously available to FSANZ (2017b) and therefore was not previously considered.

#### <span id="page-56-2"></span>**7.2.1 NTP (2022) – used by MDH (2020b), MPART (2019a), OEHHA (2022a)**

NTP (2022) conducted 28-day toxicity studies in male and female Sprague Dawley rats (n=10/dose; five doses per chemical) to compare the toxicities of seven PFAS [PFBS, PFHxS potassium salt (PFHxSK), PFOS, and four carboxylates] via gavage in deionised water with 2% Tween® 80. NTP (2022) describes the results for PFBS, PFOS and PFHxSK; a companion report describes the results for the PFAS carboxylates.

Doses for the PFHxSK (>98% purity) treated animals were 0, 0.625, 1.25, 2.5, 5 and 10 mg/kg/day for males and 0, 3.12, 6.25, 12.5, 25 and 50 mg/kg/day for females administered 7 days/week for 28 days. A PPARα agonist (Wyeth-14,643) was used for qualitative comparison to the PFAS evaluated (doses 0 to 25 mg/kg/day). The studies evaluated clinical pathology, thyroid hormones, liver expression of PPARα- and constitutive androstane receptor (CAR)-related genes, liver acyl-coenzyme A oxidase enzyme activity (males only), plasma and liver (males only) PFHxS concentrations and histopathology.

All rats administered PFHxSK survived to scheduled euthanasia and there were no significant treatment-related clinical observations or effects on body weight in males or females. There were no effects on reproductive parameter indications (e.g. sperm count and motility, cyclicity, testis and epididymis weights and histopathology). Plasma concentrations of PFHxS increased with increasing dose in males and females. Although females were administered doses five times higher than those administered to males, the female plasma concentrations were about half of the male concentrations.

In PFHxSK exposed males, the following effects were observed.

- All doses: Significant decrease in free T4, total T4 and total T3 concentrations.
- ≥ 1.25 mg/kg/day: dose-related and significant increases in absolute and relative liver weights. Decreased reticulocyte counts and decreased cholesterol.
- ≥2.5 mg/kg/day: Incidence of hepatocyte hypertrophy (mild to marked) was significantly increased. Relative right adrenal gland weight in 2.5 mg/kg/day group

 $8$  MPART (2019a) cites this study as NTP (2018) because it was referencing study tables that preceded release of the report, but MDH (2020b) and OEHHA (2022a) cite it as NTP (2019). The 2019 NTP report has since been revised and updated in 2022 (NTP 2022). Minor revisions were made in NTP (2022) from the 2019 report version, all of which are marked up and identified in Appendix F of the NTP (2022) report.



and absolute and relative weights in 5 and 10 mg/kg/day groups were significantly lower. Biological significance of the adrenal gland weight increases are not clear. Decreased triglycerides.

 10 mg/kg/day: Increased relative right kidney weight. Decreased globulin, resulting in an increase in albumin:globulin ratio.

In females, the following effects were observed.

- All doses: dose-related and significant increases in absolute and relative liver weights.
- ≥6.25 mg/kg/day: Decreased total T4.
- ≥12.5 mg/kg/day: Absolute right adrenal gland weights increased (and relative increased at 50 mg/kg/day). Biological significance of the adrenal gland weight increases are not clear. Decreased free T4.
- 50 mg/kg/day: Incidences of olfactory epithelium degeneration and olfactory epithelium hyperplasia significantly increased. There was also an increase in the incidence of olfactory epithelium inflammation suppurative in this group. These changes were primarily minimal to mild in severity.

In general, the effects in male and female rats administered PFHxSK were of lower magnitude (e.g. liver or clinical pathology findings) or not apparent compared to the effects in rats exposed to PFBS and PFOS. This corresponded, to some degree, with limited to no increases in liver *Acox1* and *Cyp* gene expression changes. Several of the effects observed in the liver were also observed in rats administered Wyeth-14,643, but effects observed outside the liver by the PFAS were not observed with Wyeth-14,643. This indicates that the liver effects are potentially not relevant to humans but relevance of effects in other organ systems cannot be discounted.

Mean plasma concentrations of PFHxS in treated male rats ranged from 66,760 to 198,300 ng/mL, whereas in females they ranged from 37,030 to 95,510 ng/mL.

Changes in thyroid hormone concentrations were observed across three PFAS (PFHxSK, PFBS and PFOS). Total T4, free T4 and total T3 largely decreased in a dose-dependent manner. The magnitude of the effect was stronger in PFBS and PFOS rats compared to PFHxSK rats. Thyroid stimulating hormone (TSH) concentrations were not consistently increased across the three chemicals or sexes in response to the lower thyroid hormone levels, nor were there any histopathological changes in the thyroid gland (e.g. hyperplasia or hypertrophy). The reason for a lack of a compensatory TSH response in the face of substantially low thyroid hormone concentrations in these PFAS studies is not clear and not consistent with a classical disruption in the hypothalamic-pituitary-thyroid axis. It has been shown that PFAS can bind to proteins including albumin and transthyretin, which are transport proteins for thyroid hormones (NTP 2022). NTP (2022) also indicated that several PFOS studies (in rats and monkeys) have shown low free T4 levels as measured by analog radioimmunoassays (RIA) (the method used in the NTP 2022 study), but no change in free T4 levels when measured by equilibrium dialysis followed by RIA (ED-RIA). NTP (2022) considered that these findings are consistent with PFOS competing with free T4 for binding to serum proteins, potentially creating a negative bias in the (competitive-binding) analog RIA method. This explanation is plausible for studies in monkeys; however it is less plausible for studies in rodents given that these species have low levels of plasma thyroid shepherding proteins such as thyroid binding globulin.

Nevertheless, decreases in total T4 and T3 were found in the rat and monkey studies with PFOS, as well as the NTP (2022) study. NTP (2022) commented that it is plausible that the decreases in total T4 and T3 are related to activation of PPARα and CAR receptors resulting



in an increase in thyroxine-UDP glucuronosyltransferase and accelerated degradation of thyroxine by the liver. This explanation is plausible in rodents. However, it is not plausible in primates where plasma clearance of T4 and T3 is primarily via diodinases and not thyroxine-UDP glucuronosyltransferase. It is noteworthy that PFHxSK had a lower response in CAR activity with a lower effect observed on thyroid hormones.

Some researchers have concluded that the administration of PFAS (PFDA and PFOS) does not cause a classical hypothyroid state (NTP 2022). Primary hypothyroidism is typically clinically characterised by increased TSH and decreased T4 (in the presence or absence of thyroid histopathology), whereas secondary hypothyroidism is typically the result of a pathological change to the pituitary. It is noted the 28-day NTP (2022) study found no significant changes to TSH levels or histopathological findings in the pituitary in PFHxSK dosed rats. It could therefore be argued that the decreased T4 and T3 observed in rats administered PFHxSK in the NTP (2022) study may not be relevant to humans.

However, in a reproductive/developmental toxicity study with PFHxS in rats (Butenhoff et al. 2009), effects on thyroid histopathology were indeed observed but these effects occurred at 4-5 times higher serum PFHxS concentrations than found to have resulted in decreased T4 and T3 levels in the NTP (2022) study. In addition, no chronic toxicity study has been conducted with PFHxS which could be used to determine whether the effects observed on thyroid hormone levels in the 28-day study are likely repeatable and relevant to humans, e.g. whether in a chronic study, the effects are repeatable and would be accompanied by changes in TSH or histopathological findings on the thyroid gland or pituitary. It is also noted that associations between PFAS exposure and thyroid hormone status have been observed in some human epidemiological studies, although the associations are not always consistent (e.g. Ballesteros et al. 2017, Boesen et al. 2020, Coperchini et al. 2021). Thus, it is concluded that potential human relevancy of the thyroid hormone changes observed in the 28-day NTP (2022) study with PFHxS cannot be discounted based on currently available information.

Because the NTP (2022) study was conducted in accordance with relevant standardised testing guidelines, evaluated a large number of endpoints, and provided serum PFHxS concentrations, it is concluded to be appropriate new information to potentially adopt/adapt for derivation of candidate guidance/guideline values for PFHxS. The candidate guidance/guideline values are summarised in **Section [7.3](#page-58-0)**.

# <span id="page-58-0"></span>**7.3 Candidate guidance/guideline values for PFHxS**

As indicated in **Section [7.2.1](#page-56-2)**, the NTP (2022) study represents new suitable information that was not previously available to FSANZ (2017b) when considering derivation of a guidance value for PFHxS, noting the uncertainty with respect to human relevancy of the effect based on currently available information and the potential conservatism in any resulting guidance value. The study has been used by three jurisdictions (MDH 2020b, MPART 2019a, OEHHA 2022a) to derive a guidance value for PFHxS, one of which (OEHHA 2022a) also met a high proportion of technical/administrative criteria for potential adoption/adaptation into the Guidelines (**Section [7.1](#page-55-1)**).

The three jurisdictions who derived a guidance value for PFHxS using the NTP (2022) study either used a POD of 32,400 ng/mL (i.e. 32.4 mg/L) which represents a lower benchmark dose for 20% reduced T4 in male rats (i.e.  $BMDL_{20}$ ) (MDH 2020b, MPART 2019a) or 28,600 ng/mL (i.e. 28.6 mg/L) which represents a lower benchmark dose for one standard deviation difference from controls (BMDL<sub>1SD</sub>) for the same effect (OEHHA 2022a). There is very little difference between these two PODs.

<span id="page-59-2"></span><span id="page-59-1"></span>To derive a human POD from the animal POD, the three jurisdictions derived a similar human clearance value / toxicokinetic adjustment factor<sup>[9](#page-59-1)</sup> (i.e. 0.000085-0.00009 L/kg-day). This resulted in very similar human equivalent dose (HED) PODs of 0.00243 to 0.00292 mg/kg/day. The jurisdictions then applied different uncertainty factors (300 or 1,000) to their HED POD (see **[Table 7-1](#page-59-0)**). The difference is primarily due to OEHHA (2022a) deciding to apply an additional uncertainty factor of 10 for the use of a sub-chronic study.

However, it is noted that a reproductive/developmental toxicity study with PFHxS in rats (Butenhoff et al. 2009) was summarised by FSANZ (2017b), in which the NOAEL (for paternal toxicity) in male rats was stated to be 3 mg/kg/day (for offspring toxicity, it was higher at 10 mg/kg/day). The serum concentration for paternal males at day 42 of the study was 128,670 ng/mL, with some effects on thyroid histopathology (hypertrophy and hyperplasia of the follicular cells) noted in males at this serum concentration. The serum concentration in males at the lower dose of 1 mg/kg/day on day 42 was 89,120 ng/mL. This indicates that histopathological effects in the thyroid are only likely to manifest at higher serum concentrations, i.e. 4-5 times higher, than decreased T4 in male rats. The half-life of PFHxS in male rats (i.e. 3.6-15.9 days or ~10 days, Benskin et al. 2009) suggests that serum PFHxS in these rats was likely at steady state. Thus, the use of an uncertainty factor for use of subchronic study (in addition to the database uncertainty factor) is unlikely to be warranted. The database uncertainty factor is likely to already account for use of a subchronic study, since the former is applied for lack of chronic toxicity studies. It is therefore suggested an overall composite uncertainty factor of 300 rather than 1,000 is likely sufficient and still provides a conservative guidance value.

With respect to the relative source contribution (RSC) factor, the current factor employed in derivation of the DWGs for PFOS, PFHxS and PFOA in the Guidelines is 0.1 (i.e. 10%) which is also the default factor for the Australian context. It is noted all jurisdictions which have derived DWGs in the literature consulted applied an RSC of 0.2 (i.e. 20%) (e.g. MDH 2020b, OEHHA 2022a) but do not provide the rationale for this. Thus, the default factor of 0.1 has been retained in calculating the potential resulting DWGs for PFHxS using these guidance values in **[Table 7-1](#page-59-0)**, noting that it yields a lower guideline value than use of an RSC of 0.2. Also presented in **[Table 7-1](#page-59-0)** is the derivation of the current Australian DWG for PFOS + PFHxS of 70 ng/L, which is based on a toxicology study for PFOS.

<span id="page-59-0"></span>

<b>Parameter</b>	NHMRC and   MDH 2022b <b>NRMMC</b> 2011, <b>FSANZ</b> 2017b, DOH 2017	<b>MPART 2019a</b>	OEHHA 2022a
Critical study	Luebker et al. 2005b	<b>NTP 2022</b>	

 $9$  i) MDH (2020b) derived the toxicokinetic adjustment factor as follows: Clearance rate = Volume of Distribution (L/kg) x (Ln2/Half-life, days); Clearance rate = 0.25 L/kg x (0.693/1935 days); Clearance rate = 0.00009 L/kg-day ii) MPART (2019a) used the same toxicokinetic adjustment factor as MDH (2020b).

iii) OEHHA (2022) derived a very similar clearance factor of 0.000085 L/kg-day.



DWG = Drinking Water Guideline; ↓ = Decreased; BW = Body weight; F0 = Parental generation; POD = Point of Departure; BMDL = Lower Benchmark Dose; HED = Human Equivalent Dose; UF $_A$  = Uncertainty factor for extrapolation from animals to humans;  $UF_H =$  Uncertainty factor for human variability;  $UF_{\text{timerframe}} =$  Uncertainty factor for use of a short-term study;  $UF_{\text{composite}} =$  Composite (i.e. total) uncertainty factor;  $UF_{\text{database}} =$ Uncertainty factor to account for the limited database of toxicological studies (e.g. no two-generation or immunotoxicity studies).

<span id="page-61-0"></span>

DWG (ng/L) = [Guidance value (ng/kg bw/day) x 70kg (adult) x 0.1 for adult]  $\div$  2 L/day for adult

The candidate PFHxS DWGs derived by adapting existing guidance values for this PFAS are 8.5 ng/L using the uncertainty factors from OEHHA (2022a) or 34 ng/L using the uncertainty factors from MDH (2020b) and MPART (2019a); as discussed in the text preceding the table, the difference between the two values is the application of an additional uncertainty factor. The value of 34 ng/L is considered to be more appropriate based on the reasons cited above the table, noting the uncertainty and likely conservatism with respect to human relevancy of the selected endpoint based on currently available information.

Assuming the recommendation in **Section [6.3](#page-53-1)** for PFOS is accepted, it is noted that, in accordance with enHealth (2016) guidance and current practice in Australia, it is considered reasonable to retain the existing guideline value of 70 ng/L as the sum of PFOS+PFHxS when evaluating concentrations in drinking water in addition to comparison of PFHxS concentrations on their own with the suggested candidate guideline value of 34 ng/L.

<span id="page-61-1"></span>In Australian distributed drinking waters, PFHxS concentrations generally may range from <2 to 5 ng/L in Queensland (QAEHS 2018a, 2018b)<sup>[10](#page-61-0)</sup>, Sydney (Sydney Water 2023) and Western Australia (WCWA 2023) which are below both candidate DWGs. However, PFOS + PFHxS concentration was measured at 90% of the current Australian DWG (i.e. ~ 60 ng/L) in one bore in a drinking water borefield supplying Esperance, Western Australia (WCWA 2019, 2020). Once this apparent PFOS/PFHxS contamination was identified, this bore was no longer used (WCWA, 2023). This indicates that compliance with the candidate DWGs may present an issue in certain circumstances. Nevertheless, due to the large uncertainty factors, likely conservatism of the selected endpoint with respect to PFHxS, and small RSC incorporated into the derivation of the candidate DWGs, PFHxS is unlikely to present a human health risk from distributed drinking water in most regions of Australia. However, there are many sites of PFAS contamination in Australia, and, if water from these contaminated sites is used as a local source of drinking water (e.g. backyard bore in rural location where distributed water is not available), PFHxS may be present at concentrations above the candidate DWG and the existing Australian DWG in these cases.

# **8.0 Discussion for PFBS**

This section provides a discussion of the strengths and limitations of the identified guidance values for PFBS for possible adoption/adaptation into the Guidelines.

## <span id="page-61-2"></span>**8.1 Potential suitability of health-based guidance values for possible adoption/adaptation**

Candidate guidance values for PFBS described in **Section [4.1](#page-17-0)** for possible adoption/adaptation in Australia have also been evaluated using the Assessment Tool provided in Appendix D in the Technical Report and already described in **Section [6.1](#page-48-2)** for PFOS.

[<sup>10</sup>](#page-61-1) Note the Queensland data are for raw water catchments.

<span id="page-62-1"></span>**[Figure 8-1](#page-62-0)** presents the percentage of criteria (combined technical and administrative criteria) met by each jurisdiction. It is evident from the figure that the highest percentage of 'must-have' and 'should-have' criteria were met by US EPA (2021c), followed by OEHHA (2021c), MPART (2019a) and then MDH (2022g).



#### <span id="page-62-0"></span>**Figure 8-1 Overall proportion of 'must-have', 'should-have' and 'may-have' technical/administrative criteria as per the Assessment Tool met by jurisdictions who have derived candidate guidance values for PFBS for possible adoption/adaptation in Australia**

## **8.2 Critical evaluation of PFBS guidance values**

As PFBS was not part of the comprehensive review undertaken by FSANZ (2017b), all guidance values sourced in the literature search for which the derivation was described were evaluated in this section. These include the following.

- 84 ng/kg/day (MDH 2022e, g) (decreased total thyroxine (T4) in rats; critical study: NTP 2022).
- 300 ng/kg/day (MPART 2019a, US EPA 2022c, k; 2021c; WSDH 2019a, 2023a, 2022b) (decreased total thyroxine (T4) in mice; critical study: Feng et al. 2017).
- <span id="page-62-2"></span> 600 ng/kg/day (OEHHA 2021d) (decreased total thyroxine (T4) in mice; critical study: Feng et al. 2017).

All jurisdictions have agreed that the most sensitive health endpoint is decreased total thyroxine (T4) in rats or mice. The critical studies underpinning the derivations of the three different guidance values are NTP  $(2022)^{11}$  $(2022)^{11}$  $(2022)^{11}$  and Feng et al.  $(2017)$ .

[<sup>11</sup>](#page-62-2) MDH (2022g) cites this study as NTP (2019). The 2019 NTP report has since been revised and updated in 2022 (NTP 2022). Minor revisions were made in NTP (2022) from the 2019 report version, all of which are marked up and identified in Appendix F of the NTP (2022) report.



### <span id="page-63-0"></span>**8.2.1 NTP (2022) – used by MDH (2022e, g)**

As described in **Section [7.2.1](#page-56-2)** for PFHxS, NTP (2022) conducted 28-day toxicity studies in male and female Sprague Dawley rats (n=10/dose; five doses per chemical) to compare the toxicities of seven PFAS [PFBS, PFHxSK, PFOS, and four carboxylates] via gavage in deionised water with 2% Tween® 80. NTP (2022) describe the results for PFBS, PFOS and PFHxSK; a companion report describes the results for the PFAS carboxylates.

Doses for the PFBS (>97% purity) treated animals were 0, 62.6, 125, 250, 500 and 1,000 mg/kg/day for both males and females administered 7 days/week for 28 days. A PPARα agonist (Wyeth-14,643) was used for qualitative comparison to the PFAS evaluated doses (0, 6.25, 12.5, or 25 mg/kg/day). The studies evaluated clinical pathology, thyroid hormones, liver expression of PPARα- and constitutive androstane receptor (CAR)-related genes, liver acyl-Coenzyme A oxidase enzyme activity (males only), plasma and liver (males only) PFHxS concentrations and histopathology.

NTP (2022) cites other studies which have shown the half-lives of PFBS after oral administration of 30 mg PFBS/kg in Sprague Dawley rats to be 4.7 and 7.4 hours in males and females, respectively; in humans, a geometric mean half-life of 25.8 days has been estimated.

In PFBS exposed males, the following effects were observed.

- All dose groups: Decreased total protein, due to decreases in globulin, which resulted in increases in albumin/globulin ratio. Decreased cholesterol. Decreased total T4, free T4 and total triiodothyronine (T3) concentrations. TSH levels were unchanged.
- ≥ 62.6 mg/kg/day: Dose-related and significant increases in relative liver weights.
- ≥125 mg/kg/day: Dose-related and significant increases in absolute and relative liver weights (except in 1,000 mg/kg/day group). Mild significant decreases in male rat erythron, characterised by decreased haematocrit, haemoglobin and erythrocyte and reticulocyte counts. Increased incidence of hepatocyte hypertrophy.
- 250 mg/kg/day: Mild significant increases in blood urea nitrogen (BUN) concentrations at this dose and 500 mg/kg/day dose, consistent with decreased water intake (i.e. dehydration). Significantly increased incidence of olfactory epithelium degeneration and olfactory epithelium hyperplasia.
- 500 mg/kg/day: Decreased absolute and relative heart and thymus weight. Increased absolute and relative kidney weights. The biological significance of these changes is not clear. Decreased triglycerides at this dose. Increased ALT, alkaline phosphatase (ALP), and aspartate aminotransferase (AST) activity. Increased sorbitol dehydrogenase (SDH) activity. Increased total bile acid concentrations. Significantly increased hepatocyte cytoplasmic alteration. Significantly increased incidence of olfactory epithelium degeneration and olfactory epithelium hyperplasia.
- 1,000 mg/kg/day: Nine of the ten rats in this group died from day 15 to day 25 and one due to a dosing accident on day 6. Seizure recorded in one male. Body weight was 17% and 19% reduced from controls at 15 and 22 days, respectively. Significantly increased hepatocyte cytoplasmic alteration. One male had hepatocyte necrosis. Increased incidence of mild to marked bone marrow hypocellularity. Significantly increased incidence of olfactory epithelium degeneration and olfactory epithelium hyperplasia. Significantly increased incidence of minimal to mild epithelium hyperplasia in forestomach. Significantly increased incidence of mild to marked thymus atrophy. One male had kidney papilla necrosis.

In females, the following effects were observed.

- All doses: Dose-related and significant increases in relative right kidney weights. Decreased total T4, free T4 and total T3 concentrations. TSH levels were unchanged.
- ≥125 mg/kg/day: Dose-related and significant increases in relative liver weight. Decreased reticulocyte counts. Significantly increased incidence of olfactory epithelium degeneration and olfactory epithelium hyperplasia (the latter from 250 mg/kg/d).
- ≥250 mg/kg/day: One rat died on day 25. Seizures recorded in one female at this dose. Dose-related and significant increases in relative and absolute liver weight. Increased total bile acid concentrations.
- 500 mg/kg/day: One rat died on day 21. Seizures recorded in two females at this dose. Decreased absolute spleen, heart and thymus weights. The biological significance of the latter changes is not clear. Decreased cholesterol at this dose. Increased ALT, ALP, and AST activity. Increased incidence of hepatocyte hypertrophy. Significantly increased hepatocyte cytoplasmic alteration.
- 1,000 mg/kg/day: Eight rats died from day 16 to 27. Seizures recorded in one female at this dose. One rat was lethargic, two had ruffled fur and two were thin. Mean body weight reduced by 14% compared to controls. Increased incidence of hepatocyte hypertrophy. Significantly increased hepatocyte cytoplasmic alteration. Increased minimal hepatocyte necrosis. Increased incidence of mild to marked bone marrow hypocellularity. Significantly increased incidence of mild to marked thymus atrophy. Significantly increased incidence of minimal to mild kidney papilla necrosis.

Except for the deaths related to the dosing accident, all other deaths were considered treatment-related but the cause undetermined. The cause of the seizures was unknown, and they were not repetitive. There was no clinical pathology interpretation in the groups administered the highest dose tested due to the high mortality in these groups.

Plasma concentrations of PFBS increased with increasing dose in both males and females, with males generally having higher (5- to 18-fold) plasma concentrations compared with females across all dose groups.

Male and female rats administered PFBS exhibited a significant increase in expression of *Acox1, Cyp4a1, Cyp2b1,* and *Cyp2b2* compared to controls, indicating significant increased PPARα and CAR activity. Males displayed a greater fold increase in PPARα-related gene expression compared to controls than females, whereas expression of CAR-related genes were more prominent in female rats.

The testicular spermatid count in the 250 mg/kg/day males was lower (10%) than the vehicle control group. When normalised to total testicular weight, counts in the 250 and 500 mg/kg/day groups were 12% and 10% lower, respectively, than the vehicle control group. These differences did not attain statistical significance, but the trend was significant. Left testis and left epididymis weights were not affected by PFBS administration. The histopathologic finding of germinal epithelium degeneration in the testis was noted in one male in the 1,000 mg/kg/day group (sperm assessments were not made in this group due to early mortality). Serum testosterone levels assessed at necropsy in dosed males were similar to the vehicle control group level. Females administered 250 or 500 mg/kg/day PFBS displayed alteration in the oestrous cycle (extended diestrus in the 250 mg/kg/day females, irregular or not cycling in the 500 mg/kg/day females).

Several of the effects observed in the liver were also observed in rats administered Wyeth-14,643, but effects observed outside the liver with PFAS administration were not observed with Wyeth-14,643. This indicates that the liver effects are potentially not relevant to humans but relevance of effects in other organ systems cannot be discounted.



Mean plasma concentrations of PFBS in treated male rats ranged from 2,222 to 43,160 ng/mL (the latter in the 500 mg/kg/day dose group), whereas in females they ranged from 154 to 24,455 ng/mL. However, it is unclear from the study at which time-point postadministration of the final dose these plasma concentrations were measured. This is important for PFBS; due to the relatively short half-life of PFBS in rats (i.e. 4.7-7.4 hours), depending on when samples were collected for analysis, plasma concentrations shortly after administration may have been 2-3 times higher than reported in the study.

MDH (2022e, g) considered the decreased total T4 observed at all doses in female rats to be the critical adverse endpoint and derived a POD as a  $BMDL_{1SD}$  of 6.97 mg/kg/day for this effect. They derived a chemical-specific toxicokinetic adjustment factor of 0.0012 representing the difference in half-lives between female rats (1.3 hours) and humans (1050 hours), i.e. 1.3 h  $\div$  1050 h = 0.0012. MDH (2022e, g) then derived a HED by multiplying the POD by the toxicokinetic adjustment factor  $(6.97 \text{ mg/kg/day} \times 0.0012 = 0.0084 \text{ mg/kg/day})$ and dividing this dose by an uncertainty factor of 100 (3x for interspecies differences in toxicodynamics; 10x for intraspecies variability; 3x for database uncertainty due to lack of immunotoxicity, developmental neurotoxicity studies, or 2-generation toxicity study) resulting in a TRV of 84 ng/kg/day. It is noted that if the half-lives cited in the NTP (2022) study (7.4  $\overline{h}$ in female rats, 619 h in humans) were used instead, the toxicokinetic adjustment factor would change to 0.012 (an order of magnitude difference), and if all other considerations remained the same, this would change the TRV to 833 ng/kg/day (one order of magnitude higher). This highlights the sensitivity of the value of the TRV to the appropriate half-life information.

Similar to the discussion for PFHxS in **Section [7.2.1](#page-56-2)** with respect to this study, the decreased T4 and T3 observed in the NTP (2022) study in rats administered PFBS was not accompanied by increased TSH or thyroid histopathological findings. This indicates there is uncertainty with respect to the human relevancy of the effect based on currently available information. Nevertheless, it is noted that a developmental/reproductive toxicity study in mice by Feng et al. (2017), described in **Section [8.2.2](#page-65-0)** below, also found decreased T3 and T4 levels at postnatal day 30 which were accompanied by slight but statistically increased serum TSH. In addition, the human epidemiological literature has found associations between PFAS exposure and thyroid hormone changes (see **Section [7.2.1](#page-56-2)**), albeit these associations were not always consistent. As there is a lack of chronic toxicity studies with PFBS (similar to PFHxS), and the Feng et al. (2017) study found increased TSH accompanied the decreased T3 and T4 levels, it is concluded that the potential human relevancy of this effect for PFBS cannot be discounted based on currently available information.

It is noted that OEHHA (2021d) did not use the NTP (2022) study in rats for their POD when deriving a guidance value because of the large toxicokinetic differences between female rats and humans, and the uncertainty around the utility of the rat model for effects in humans of maternal thyroid hormone disruption of foetal development.

Because the NTP (2022) study was conducted in accordance with relevant standardised testing guidelines and evaluated a large number of endpoints, it is concluded to be appropriate information to potentially adopt/adapt for derivation of candidate guidance/guideline values for PFBS. The candidate guidance/guideline values are summarised in **Section [8.3](#page-67-0)**.

#### <span id="page-65-0"></span>**8.2.2 Feng et al. (2017) – used by MPART (2019a), US EPA (2022c, k; 2021c), WSDH (2019a, 2023a, 2022b), OEHHA (2021d)**

Feng et al. (2017) investigated the influence of gavage exposure to K<sup>+</sup> PFBS (98% purity) (0, 50, 200 or 500 mg/kg/day) in 0.1% carboxymethylcellulose during gestation days 1 to 20 on perinatal growth and development, pubertal onset, and reproductive and thyroid function in



female ICR mice. On postnatal day (PND) 21, all offspring were weaned. Female offspring were transferred to other cages (2-4 per cage). Thirty dams in each dose group were randomly assigned to one of the following three experimental groups: i) group 1, in which perinatal survival and growth, pubertal onset, and ovarian and uterine development were sequentially examined in the same cohorts (50 offspring/10 dams); ii) group 2, in which hypothalamic–pituitary–gonadal hormone and hypothalamic–pituitary–thyroid hormone levels were measured in PND1 offspring ( $n = 30$ ), PND30 offspring ( $n = 10$ ), and PND60 offspring ( $n = 10$ ) obtained from 10 dams; and iii) group 3, in which the levels of serum PFBS were measured ( $n = 10$  dams).

The weight gain of the dams was not different between the different treatment groups. Dams did not exhibit foetal loss or abnormal behaviour during the administration of PFBS.

Number of neonatal PFBS-offspring were not significantly different from that in the control group. All offspring appeared to be active and survived until adulthood. The potentially treatment-related findings in the study were as follows.

- $\bullet$   $\geq$  200 mg/kg/day:
	- o Body weights of PND1 female PFBS-offspring were significantly lower compared to controls. These offspring remained underweight throughout weaning, pubertal and adult periods.
	- o Slight but statistically significant delay (approximately 1.5-2 days) in eye opening, delay in vaginal opening, and delay in first oestrous (of up to 5 days) observed in treated offspring compared with control offspring (p<0.01). Size of the ovaries of PND60 treated offspring were smaller than those of controls and relative weights were lower (p<0.05). PFBS treated offspring at these doses exhibited fewer primordial follicles, primary follicles, secondary follicles, early antral follicles, antral follicles and pre-ovulatory follicles, as well as fewer corpora lutea (p<0.05) than controls at diestrus. PND40-60 offspring in these dose groups exhibited a prolongation of diestrus compared with controls (p<0.05) with reduced serum E2 levels (p<0.05) in PND30 and PND60 offspring, a slight increase in luteinising hormone level in the PND30 offspring only, but no difference in hypothalamic gonadotropin-releasing hormone compared to controls.
	- PND1, PND30 and PND60 offspring in these groups exhibited significantly reduced serum total T3 and T4 levels compared with controls, with the reduction in total T4 lower at PND60 (23%) compared with PND30 (42%). In addition, PND30 offspring in these dose groups showed slight but statistically significant elevations in serum thyroid stimulating hormone (TSH) (p<0.05).
	- o PFBS treated dams in these dose groups exhibited statistically significantly reduced total T4 ( $p<0.05$ ), total T3 ( $p<0.05$ ) and free T4 ( $p<0.05$ ), as well as increased TSH (p<0.05).

The information summarised above indicates a PFBS dose of 50 mg/kg/day was the NOAEL in this study.

Serum PFBS in treated pregnant mice (collected 12 hours after the last administered dose) were  $1.73 \pm 0.65$  ng/mL,  $74.01 \pm 22.52$  ng/mL,  $332.41 \pm 53.04$  ng/mL and  $720.86 \pm 98.4$ ng/mL in the 0, 50, 200, and 500 mg/kg/day groups, respectively. Half-lives of PFBS in male and female CD-1 mice have been reported at 5.8 h in males and 4.5 h in females (Lau et al. 2020) and in humans 619 h (as per NTP 2022). Since serum collection in the Feng et al. (2017) study occurred 12 hours after the last administered dose, serum concentrations in dams are likely to have been 2.7x higher (i.e. ~200 ng/mL at NOAEL dose of 50 mg/kg/day; ~900 ng/mL at LOAEL dose of 200 mg/kg/day) directly after administration of the last dose.



<span id="page-67-1"></span>The data from the Feng et al. (2017) study was used to derive TRVs by various jurisdictions as follows.

- MPART (2019a) considered the critical effect to be decreased serum total T4 in PND1 mice and derived a BMDL<sub>20</sub> of 28.19 mg/kg/day for this effect. They divided this BMDL<sub>20</sub> by a toxicokinetic adjustment factor of 316 (i.e. human serum half-life of 665 h  $\div$  female mouse serum half-life of 2.1 h) to derive a HED POD of 0.0892 mg/kg/day. This was divided by an uncertainty factor of 300 (10x for human variability; 3x for interspecies variability in toxicodynamics, 10x for database deficiencies due to lack of neurodevelopmental, immunotoxicological and chronic studies) to derive a TRV of 300 ng/kg/day. WSDH (2019a, 2023a) derived the same TRV in the same manner as MPART (2019a).
- OEHHA (2021d) considered both the NTP (2022) and Feng et al. (2017) studies for deriving a TRV but decided against using the NTP (2022) study because of the large toxicokinetic differences between female rats and humans, and uncertainty around the utility of the rat model for effects in humans of maternal thyroid hormone disruption on foetal development. They derived a similar POD to MPART (2019a) but expressed it as a BMDL<sub>1SD</sub> of 22.2 mg/kg/day. They adjusted this by a clearance factor of 345 [Ratio of animal to human clearance =  $(0.056 \text{ L/kg/h} \times 1000 \text{ mL/L} \times 24$ h/day)  $\div$  3.9 mL/kg/day = 3451 to derive a HED POD of 0.064 mg/kg/day. They applied an uncertainty factor of 100 (√10 for interspecies differences in toxicodynamics; 10x for human variability; √10 for database deficiencies) to derive a TRV of 600 ng/kg/day (rounded).
- <span id="page-67-2"></span> US EPA (2021c, 2022c, 2022k) also agreed with the critical endpoint of decreased total T4 in newborn mice in the Feng et al. (2017) study. They derived a POD as a lower benchmark dose for half a standard deviation difference from controls (BMDL<sub>0.5SD</sub>) of  $\sim$ 22.1 mg/kg/day<sup>[12](#page-67-1)</sup> and applied a toxicokinetic adjustment factor of 0.0043 (i.e. 4.5 h in female mice  $\div$  1050 h in humans) to derive a HED POD of 0.095 mg/kg/day. Note if the human half-life cited in NTP (2022) of 619 h was used, this factor would be 0.0073 (~1.7x difference). This was divided by an uncertainty factor of 300 (3x for interspecies toxicodynamics; 10x for human variability; 10x for database uncertainties) to derive a TRV of 320 ng/kg/day.

The Feng et al. (2017) study was peer reviewed, appears to have been conducted appropriately and evaluated relatively sensitive endpoints of interest (female reproductive performance and developmental effects); it is concluded to be appropriate information to potentially adopt/adapt for derivation of candidate guidance/guideline values for PFBS. The candidate guidance/guideline values are summarised in **Section [8.3](#page-67-0)**.

# <span id="page-67-0"></span>**8.3 Candidate guidance/guideline values for PFBS**

As indicated in **Section [8.2.1](#page-63-0)** and **[8.2.2](#page-65-0)**, both the NTP (2022) and the Feng et al. (2017) studies represent suitable information for potential guidance value derivation for PFBS, noting the uncertainty with respect to human relevancy of the effect based on currently available information and the potential conservatism in any resulting guidance value. The studies have been used by five jurisdictions (MDH 2022e, g; MPART 2019a; OEHHA 2021d;

<sup>&</sup>lt;sup>[12](#page-67-2)</sup> Note SLR has estimated this  $BMDL<sub>0.5SD</sub>$  from the information in the US EPA (2021c) report by back-calculating from the POD HED cited in the report (0.095 mg/kg/day) and dividing by the dosimetric adjustment factor derived for female mice compared with humans (see Table 8 in US EPA 2021c). The adjustment factor was 0.0043 (i.e. 4.5 h in female mice ÷ 1050 h in humans). Note if the human half-life cited in NTP (2022) of 619 h was used, this factor would be 0.0073 (~1.7x difference).



US EPA 2021c, 2022c, 2022k; WSDH 2019a, 2023a) to derive a guidance value for PFBS, two of which (OEHHA 2021d; US EPA 2021c, 2002c, 2022k) also met a relatively high proportion of technical/administrative criteria for potential adoption/adaptation into the Guidelines (**Section [8.1](#page-61-2)**).

The four jurisdictions who derived a guidance value for PFBS using the Feng et al. (2017) study used very similar PODs ranging from 22.1 to 28.19 mg/kg/day for the same critical effect. The jurisdiction that derived a guidance value using the NTP (2022) study derived a different POD of 6.97 mg/kg/day.

To derive a human POD from the animal POD, the various jurisdictions derived human toxicokinetic adjustment factors for the difference between human half-lives and rat or mouse half-lives, depending on the study species; the factors (as the ratio of human to animal half-life) ranged from 233 to 345 for mice and 808 for the rat study. This resulted in similar HED PODs of 0.064 to 0.095 mg/kg/day for the mouse and 0.0084 mg/kg/day for the rat study; although it is noted if the half-lives cited in NTP (2022) were used the latter HED POD would be an order of magnitude higher (0.084 mg/kg/day) and fall within the range of the mouse HED PODs. The jurisdictions then applied different uncertainty factors (100 or 300) to their HED POD (see **[Table 8-1](#page-68-0)**). The difference is due to application of an uncertainty factor of 3 or 10 for database uncertainties.

With respect to the relative source contribution (RSC) factor, the current factor employed in derivation of the DWGs for PFOS, PFHxS and PFOA in the Guidelines is 0.1 (i.e. 10%) which is also the default factor for the Australian context. It is noted all jurisdictions which have derived DWGs in the literature consulted applied an RSC of 0.2 (i.e. 20%) (e.g. MPART 2019a, OEHHA 2021d, US EPA 2022c, k) but do not provide the rationale for this. Thus, the default factor of 0.1 has been retained in calculating the potential resulting DWGs for PFBS using these guidance values in **[Table 8-1](#page-68-0)**, noting that it yields a lower guideline value than use of an RSC of 0.2.



#### <span id="page-68-0"></span>**Table 8-1 Potential drinking water guideline values (ng/L) resulting from adaptation of PFBS guidance values from different jurisdictions based on two critical studies**



DWG = Drinking Water Guideline; BMDL = Lower Benchmark Dose; HED = Human Equivalent Dose; GD = Gestation Day; PND = Postnatal Day; UF<sub>A</sub> = Uncertainty factor for extrapolation from animals to humans; UF<sub>H</sub> = Uncertainty factor for human variability;  $UF_{\text{imeframe}} =$  Uncertainty factor for use of a short-term study;  $UF_{\text{composite}} =$  Composite (i.e. total) uncertainty factor;  $UF_{\text{database}} =$  Uncertainty factor to account for the limited database of toxicological studies (e.g. no two-generation or immunotoxicity studies).

(1) Adaptation of guidance value has been undertaken using the default assumptions for derivation of DWGs in Australia using the following equation as outlined in NHMRC (2021):

DWG (ng/L) = [Guidance value (ng/kg bw/day) x 70kg (adult) x 0.1 for adult]  $\div$  2 L/day for adult

(2) As highlighted in the text in **Section [8.2.1](#page-63-0)**, the toxicokinetic adjustment factor is very sensitive to the input half-lives assumed for female rats and humans. If the half-lives cited by NTP (2022) of 7.4 h in female rats and 619 h in humans are used instead, the adjustment factor would decrease by a factor of 10, thereby increasing the POD HED and resulting TRV by a factor of 10. The values that would result from using the half-lives cited by NTP (2022) are provided in brackets.

The candidate PFBS DWGs derived by adapting existing guidance values for this PFAS are 302 (or 2,939) ng/L using the rat toxicology study (NTP 2022) or range from 1,041 to 2,252 ng/L using the mouse toxicology study by Feng et al. (2017). The guideline values resulting from adapting the TRV from the rat study (using the half-lives cited in NTP 2022) and the TRVs from the mouse toxicology study are within a factor of three (ranging from 1,041 to 2,939 ng/L) and are considered most applicable within the Australian context. It is reiterated that the endpoint on which these guidance values are based is of uncertain human relevance based on currently available information and therefore the resulting candidate guideline values are conservative.

In Queensland raw water catchments, PFBS concentrations have been recorded up to 2.2 ng/L (QAEHS 2018a, 2018b). There are few PFBS data in drinking water elsewhere in Australia. Based on the limited data available, it appears that PFBS concentrations in distributed drinking water in Australia are markedly lower than any of the candidate DWGs, suggesting PFBS is unlikely to present a human health risk from distributed drinking water in Australia. However, there are many sites of PFAS contamination in Australia, and, if water from these contaminated sites is used as a local source of drinking water (e.g. backyard bore in rural location where distributed water is not available), PFBS may be present at concentrations above the candidate DWGs in these cases.

# **9.0 Discussion for PFOA**

This section provides a discussion of the strengths and limitations of the identified guidance values for PFOA for possible adoption/adaptation into the Guidelines. Critical evaluation was focused on those guidance values derived using underpinning studies not previously considered / evaluated by FSANZ (2017b).

## **9.1 Potential suitability of health-based guidance values for possible adoption/adaptation**

Candidate guidance values for PFOA described in **Section [4.1](#page-17-0)** for possible adoption/adaptation in Australia have also been evaluated using the Assessment Tool provided in Appendix D in the Technical Report and already described in **Section [6.1](#page-48-2)** for PFOS.

**[Figure 9-1](#page-70-0)** presents the percentage of criteria (combined technical and administrative criteria) met by each jurisdiction. It is evident from the figure that several publications met similar percentages of criteria, with ATSDR (2021a), EFSA (2020a), FSANZ (2017b), NJDEP (2019a), OEHHA (2023a), and US EPA (2022c, d; 2021a) all meeting relatively high (i.e. ~>60%) proportions of 'must-have' and 'should-have' criteria.

Other jurisdictions (HC 2018b, MDH 2022f, OEHHA 2019a, MPART 2019a) met lower proportions of criteria, indicating these guidance documents potentially do not conform with modern methods of undertaking systematic reviews.



#### <span id="page-70-0"></span>**Figure 9-1 Overall proportion of 'must-have', 'should-have' and 'may-have' technical/administrative criteria as per the Assessment Tool met by jurisdictions who have derived candidate guidance values for PFOA for possible adoption/adaptation in Australia**

# **9.2 Critical evaluation of PFOA guidance values**

For PFOA, the guidance values identified in the literature review that based their derivations on underpinning studies not previously considered / cited in the comprehensive review



<span id="page-71-0"></span>undertaken by FSANZ (2017b) are the following. The discussion in this section therefore focuses on the relevant studies underpinning these guidance values.

- EFSA (2020a) who derived a guidance value for ∑PFOA, PFNA, PFHxS and PFOS of 0.63 ng/kg bw/day (TWI = 4.4 ng/kg bw per week) for decreased antibody titre for specific vaccines. The critical study underpinning this guidance value (Abraham et al. 2020) has already been critically evaluated in **Section [6.2.1](#page-50-0)**. This study was not available to FSANZ at the time of their 2017 review but was reviewed later by FSANZ (2021).
- US EPA (2022c, d; 2021a) who derived a DRAFT guidance value of 0.0015 ng/kg/day for PFOA based on decreased antibody titre following tetanus vaccination in 7-year old children. The critical studies underpinning this guidance value have already been critically evaluated either by FSANZ (2017b) (in the case of Grandjean et al. 2012), FSANZ (2021), or in **Section [6.2.2](#page-52-2)** (in the case of Budtz-Jørgensen and Grandjean 2018).
- <span id="page-71-1"></span>OEHHA (2019a) derived a non-cancer<sup>[13](#page-71-0)</sup> guidance value of 0.45 ng/kg/day for liver toxicity (and oxidative DNA damage, changes in mitochondrial membrane potential) in female mice. The critical study underpinning this guidance value is Li et al. (2017) and was not previously available to FSANZ (2017b). Therefore, this study has been critically evaluated in **Section [9.2.1](#page-72-0)**.
- In a later document, OEHHA (2023a) derived a non-cancer guidance value of 0.87 ng/kg/day for increased risk of elevated alanine aminotransferase (ALT) in humans. The critical study underpinning this guidance value is Gallo et al. (2012) which does not appear to have been evaluated by FSANZ (2017b), as assumed by a lack of its citation in the review. Therefore, this study has been critically evaluated in **Section [9.2.2](#page-73-0)**.
- NJDEP (2019a) derived a guidance value of 2 ng/kg/day for increased liver weight in male mice. The critical study underpinning this guidance value is Loveless et al. (2006), which was also not cited in the FSANZ (2017b) review. Therefore, this study has been critically evaluated in **Section [9.2.3](#page-75-0)**.
- ATSDR (2021a; adopted by WSDH 2019a, 2022b, 2023a) derived a guidance value of 3 ng/kg/day for skeletal alterations in adult mouse offspring. The critical study underpinning this guidance value is Koskela et al. (2016) which does not appear to have been previously evaluated by FSANZ (2017b). Therefore, this study has been critically evaluated in **Section [9.2.4](#page-76-0)**.
- MPART (2019a) derived a guidance value of 3.9 ng/kg/day for developmental delays (decreased number of inactive periods, altered novelty induced activity and skeletal alteration such as bone morphology and bone cell differentiation in the femurs and tibias) of mice. The critical studies underpinning this guidance value are Koskela et al. (2016) and Onishchenko et al. (2011), neither of which appear to

[<sup>13</sup>](#page-71-1) As indicated in the tables of the Technical Report and tables in **Section 4.1** of this report, the cancer-based guidance values derived by some jurisdictions were not considered to be applicable to an Australian context, as they use low-dose linear extrapolation as a policy decision in their derivations; Australia's science policy is to only undertake low-dose linear extrapolation for carcinogens acting through a mutagenic mode of action. As there is agreement in the various jurisdictional reviews sourced for this investigation that PFAS are not regarded as being directly mutagenic (see data extraction tables in Technical Report), the guidance values derived for cancer endpoints by low-dose linear extrapolation are not considered applicable to the Australian context and have not been reviewed / critiqued further.


<span id="page-72-1"></span>have been previously evaluated by FSANZ (2017b). Therefore, these studies have been critically evaluated in **Section [9.2.4](#page-76-0)** and **[9.2.5](#page-78-0)**.

In addition, due to there being several differing candidate guideline values for PFOA, their overall confidence was assigned as being 'High', 'Moderate', 'Low', or 'Very low' based on expert judgement; this was based on an assessment of underpinning critical study quality, with rationale for the rating provided in the critical evaluation discussions of the respective underpinning study (see **Sections [9.2.1](#page-72-0)** to **[9.2.5](#page-78-0)**). This was done to provide the Committee with more information to enable comparison of the different candidate guideline value options against the current Australian guideline value to facilitate an informed decision of whether revision of the existing Australian guideline value is warranted or not.

At the request of NHMRC and the Committee, the critical study underpinning the existing Australian guidance / guideline value for PFOA (i.e. Lau et al. 2006) was also assessed for its overall confidence (see **Section [9.2.6](#page-79-0)**).

#### <span id="page-72-0"></span>**9.2.1 Li et al. (2017) – used by OEHHA (2019a)**

Li et al. (2017) divided 6-week old Balb/c mice into groups (30/sex/group) and administered each mouse PFOA (98% pure) orally via gavage at doses of 0, 0.05, 0.5, or 2.5 mg/kg/day in corn oil for 28 days. After 28-days of exposure, mice were sacrificed to collect liver and blood samples. Liver and serum samples of 10 mice from each treatment were pooled and homogenised and analysed for PFOA. Liver samples were examined for histology and proteomic change using isobaric tags for relative and absolute quantitation (iTRAQ) and Western Blotting.

In PFOA exposed males, the following effects were observed.

- ≥ 0.5 mg/kg/day: Significantly increased liver weight. Increased incidence of hepatocellular hypertrophy.
- 2.5 mg/kg/day: Decreased body weight gain at 21 days compared to controls. Signs of apoptosis of liver cells.

In females, the following effects were observed.

- All doses: Increased oxidative DNA damage, changes in mitochondrial membrane potential, and increased biomarkers of apoptosis in the liver.
- ≥ 0.5 mg/kg/day: Significantly increased liver weight. Increased incidence of hepatocellular hypertrophy.
- 2.5 mg/kg/day: Signs of apoptosis of liver cells.

Proteomic profiling revealed that reactive oxygen species (ROS) hyper-generation induced by suppression of Complex I was the major pathway to induce apoptosis in female mice at 0.05 mg/kg/day, while PPARα-activation (a mechanism considered not to be relevant to humans) was the mechanism for male mice. A recent review (Corton et al. 2018) indicates that there are a number of modulating factors, such as increased oxidative stress, that potentially alter the ability of PPARα activators to increase rodent liver effects and cancer while not being key events themselves. This indicates the potential that the effects on apoptosis observed in male and female mice by Li et al. (2017) may not be relevant to humans. FSANZ (2017b) concluded in their review that PFOA is known to cause peroxisome proliferation, leading to hepatocellular hypertrophy and increased liver weight, particularly in rodents. Although some liver pathology was seen in some animal studies with PFOA, and there is some evidence of effects of PFOA on the liver that are not mediated by PPARα receptors, it is difficult to separate the effects of PPARα activation from direct effects of PFOA on the liver (FSANZ 2017b). OEHHA (2019a, 2023a) identified a LOAEL of 0.05 mg/kg/day for changes in mitochondrial membrane potential (indicative of mitochondrial



<span id="page-73-4"></span><span id="page-73-2"></span><span id="page-73-0"></span>dysfunction), increases in biomarkers of apoptosis (caspase-9 and p53) and increased oxidative DNA damage. It is arguable whether these effects, on their own, can be considered adverse therefore the lowest dose could also be regarded as a NOAEL.

PFOA concentrations in liver and serum increased with PFOA dose, with PFOA concentrations generally higher in liver than serum. The mean serum PFOA concentrations in mice in the 0.05, 0.5 and 2.5 mg/kg/day dose groups were in females / males, respectively: 970 / 1,200 ng/mL; 2,700 / 5,900 ng/mL; 9,500 / 13,400 ng/mL (Li et al. 2017, OEHHA 2023a).

<span id="page-73-6"></span><span id="page-73-1"></span>OEHHA (2019a) derived a guidance value using what they considered to be a serum LOAEL of 970 ng/mL. They applied an uncertainty factor of 300 (3x for interspecies extrapolation of toxicodynamics, 10x for human variability, 3x for use of a LOAEL, 3x for database uncertainties due to potential for developmental toxicity at the POD)<sup>[14](#page-73-0)</sup> to derive a target human serum level of 3.2 ng/mL. This was converted to a HED of 0.45 ng/kg/dav  $10.0032$ mg/L x 1.4 x 10<sup>-4</sup> L/kg/day x 10<sup>6</sup> ng/mg]. It is noted no such database uncertainty factor was considered to be required by FSANZ (2017b) when deriving a guidance value for PFOA, thus this uncertainty factor would not be considered relevant in the Australian context. In addition, if the POD were considered a NOAEL instead of a LOAEL (as the data suggest), the TRV (without the LOAEL and database uncertainty factors) would be 4.5 ng/kg/day. As indicated above, it is also arguable whether the effects observed on the liver in this study are relevant to humans, particularly as humans are potentially refractory to these types of effects.

Li et al. (2017) was a study focusing on molecular mechanisms of PFOA-induced hepatocyte apoptosis in mice, therefore it did not follow standardised protocols for toxicity experiments. Nevertheless, it provided serum PFOA concentrations, and examined effects on the liver and therefore could be used in a weight of evidence approach for derivation of candidate guidance/guideline values for PFOA. However, it is arguable whether the effects observed at the lowest dose (0.05 mg/kg/day) in female mice can be considered adverse and whether humans may be refractory to liver effects due to PFOA exposure, thus relatively low confidence is assigned to the candidate guidance/guideline value derived using the Li et al. (2017) study. The candidate guidance/guideline values are summarised in **Section [9.3](#page-80-0)**.

#### **9.2.2 Gallo et al. (2012) – used by OEHHA (2023a)**

<span id="page-73-5"></span><span id="page-73-3"></span>In a cross-sectional study, Gallo et al. (2012) analysed data for  $46,452$  adults<sup>[15](#page-73-2)</sup> from the C8 Health Project.<sup>[16](#page-73-4)</sup> They fitted linear regression models for natural log (In)-transformed values of alanine transaminase (ALT), γ-glutamyltransferase (GGT) and direct bilirubin on PFOA, PFOS, and potential confounders (age, physical activity, body mass index, average household income, educational level, race, alcohol consumption, and cigarette smoking).

[<sup>16</sup>](#page-73-5) From 1950 through 2005, a chemical plant in the Mid-Ohio Valley, West Virginia (USA), emitted PFOA into the surrounding environment. In 2001, a group of residents filed a class action lawsuit alleging health damage from the drinking water supplies drawing on PFOA-contaminated groundwater. Part of the pre-trial settlement of the class action lawsuit included a baseline survey, the C8 Health Project, conducted in 2005-2006, that gathered data from >69,000 persons from six contaminated water districts surrounding the plant. Gallo et al. (2012) used these data to examine the cross-sectional association between serum PFOA and PFOS concentrations and markers of liver function in adults.



<sup>&</sup>lt;sup>[14](#page-73-1)</sup> It appears OEHHA (2019a) have rounded up the uncertainty factor of 270 to 300.

[<sup>15</sup>](#page-73-3) 56,554 adults (≥18 years of age) were considered for the analysis, and a total of 46,452 of those adults (82.1%) were included in the final analysis after exclusion of subjects with missing data on socioeconomic status, alcohol consumption, or cigarette smoking and other potential confounding variables or without PFAS or liver enzyme measurements.

<span id="page-74-0"></span>Logistic regression models were fitted comparing deciles of PFOA or PFOS concentrations in relation to biomarker levels. A multilevel analysis was also undertaken comparing the association of PFOA with liver biomarkers at the individual level within water districts to that at the population level between water districts.

PFOA and PFOS were associated with all potential confounders considered. Ln-transformed values of ALT were significantly associated with ln-PFOA and ln-PFOS in linear regression models [fully adjusted (model 3) coefficient: PFOA, 0.022; 95% confidence interval (CI): 0.018, 0.025; PFOS, 0.020; 95% CI: 0.014, 0.026) with a partial R<sup>2</sup> greater for the association with PFOA (0.002) than for PFOS (<0.001). A steady increase in fitted levels of ALT per decile in PFOA or PFOS serum concentrations was found, with a possible levelling off effect after approximately 30 ng/mL (when ALT was ~22.5 IU/L). This positive association was also observed in logistic regression models with a steady increase in odds ratio (OR) estimates across deciles of both PFOA and PFOS concentrations ( $p = <0.001$ ) and a significant OR for both ln-unit of PFOA (OR = 1.1; 95% CI 1.07, 1.13) and ln-unit of PFOS (OR = 1.13; 95% CI 1.07, 1.18).

Fitted values of GGT by deciles of PFOA showed an apparent positive association although it was less clear than that for ALT. The suggested association was not confirmed in the logistic regression model, in which no trend across deciles was observed (p=0.213) or for the linear ln-units of PFOA values ( $OR = 1.01$ ; 95% CI 0.99, 1.04).

For direct bilirubin, there was a suggestion of an inverse U-shaped relationship with PFOA, with increasing levels of bilirubin per increasing levels of PFOA at low PFOA levels and decreasing bilirubin levels for concentrations of PFOA above about 40 ng/mL. The linear regression relationship failed to show any association in the adjusted model.

Multilevel analysis was restricted to subjects living in water districts supplied by contaminated water (n=26,777) and excluding those with private wells. There was a significant difference between the between- and within- district components ALT and direct bilirubin; however, each outcome showed different patterns. The between- water- district regression coefficient from linear regression of ln-PFOA and ALT (0.010; 95% CI: –0.001, 0.020) was lower than the within- water- district coefficient (0.027; 95% CI: 0.022, 0.031). However, both coefficients were significant or borderline significant, in the same direction, and consistent with a positive association between ALT and PFOA levels.

<span id="page-74-1"></span>The authors found significance of associations of ALT outside the 'normal range' used in the study (i.e. cutoffs of 45 IU/L in men and 34 IU/L in women) $17$ , however only a small proportion of people had ALT values outside the selected 'normal range', making the observed values difficult to interpret in terms of a true adverse effect. Gallo et al. (2012) state that it is not clear if this small increase in ALT levels can lead to clinically diagnosable conditions or if this effect is reversible. Gallo et al. (2012) also state that data from their study cannot be directly used for estimating single-subject damage in relation to PFAS exposure. It is also noted that the reference ranges for ALT can vary depending on the laboratory. For example, Mayo Clinic (2023) cite a standard reference range for ALT of 7 to 55 IU/L. Regardless of the reference range used, the positive association observed for PFOA and ALT appears to level off within the reference range of ALT (i.e. at ~22.4 IU/L), raising uncertainty with respect to the clinical relevance of the association observed. It therefore becomes arguable whether a cross-sectional study result (recognising it was well conducted and for a relatively large population) for a positive association of serum PFOA with a biomarker of a potential effect should be used as the basis of deriving a health-based guidance value.

<sup>&</sup>lt;sup>[17](#page-74-1)</sup> These values are clinically based reference levels used by the International Federation of Clinical Chemistry and Laboratory Medicine and were approximately the 90<sup>th</sup> percentile of all ALT values in the study.



<span id="page-75-0"></span>The study authors indicate the main limitation of the study is its cross-sectional design, which makes causal inference difficult. However, the consistency of findings with other literature, in particular for the association with ALT, reinforces the hypothesis of a true association (Gallo et al. 2012).

Based on the above evaluation, it is concluded the OEHHA (2023a) guidance value based on the Gallo et al. (2012) study is not suitable for adoption/adaptation in the Australian context and it has not been included in the candidate guidance/guideline value derivation for PFOA in **Section [9.3](#page-80-0)**.

#### **9.2.3 Loveless et al. (2006) – used by NJDEP (2019a)**

Loveless et al. (2006) compared the toxicity of linear ammonium perfluorooctanoate (APFO) with that of 80% linear 20% branched chain APFO (97.99% pure), and a 100% branched form in both rats and mice. The description of the study focuses on the results in mice, as these were used by NJDEP (2019a) for derivation of a guidance value for PFOA. Male Crl:CD-1(ICR)BR mice (10/group) were gavage dosed in NANOpure® water with 0, 0.3, 1, 3, 10, or 30 mg/kg/day of the different APFO form for 14 days. The study was conducted in compliance with US EPA TSCA (40 CFR part 792) Good Laboratory Practice Standards. The study monitored for body weight, clinical signs, mortality, food consumption, clinical pathology (serum lipid parameters), liver and kidney weight, and hepatic ß-oxidation analysis (a measure of peroxisome proliferation).

There were no adverse clinical signs of toxicity observed in any treated mice. One mouse dosed with 30 mg/kg/day linear APFO died during the study with cause of death being undetermined. Mean body weights were significantly lower compared to controls following 7 and 13 days of dosing with 30 mg/kg/day of linear/branched APFO or linear APFO. Mean body weights of control mice increased 1-2 g over the course of the study, whereas in groups treated with ≥ 10 mg/kg/day linear/branched APFO or linear APFO body weights decreased between 1-6g. Treatment-related increases in liver weights decreased the apparent magnitude of body weight effects.

All three forms decreased total and high-density lipoprotein (HDL) cholesterol but triglycerides (Tg) were increased at lower doses. The LOEL was 0.3 mg/kg/day for all of the APFO forms, based on increased relative liver weight, peroxisomal ß-oxidation activity (and increased Tg for linear/branched material). Absolute liver weight was also significantly increased with ≥ 3 mg/kg/day. Serum PFOA (collected approximately 24 hours after the last dose) at the LOEL of 0.3 mg/kg/day ranged from 10,000-14,000 ng/L.

It is noted that an increase in liver weight, in the absence of histopathological findings, may be indicative of an adaptive response (ATSDR 2021a). Nevertheless, the effect has been noted to occur in other animal studies with PFOA; as it was accompanied by peroxisome proliferation, humans may be less susceptible to the effect although NJDEP (2019a) notes that similar increases in liver weight were observed in a 90-day study in cynomolgus monkeys at comparable serum levels to those observed in mice. NJDEP (2019a) also notes that increases in liver weight and other types of hepatic toxicity occur through both PPARα dependent and independent modes of action and are considered relevant to humans. ATSDR (2021a) did not consider the liver effects (increase in liver weight, hepatocellular hypertrophy, alterations in serum lipids in the absence of other degenerative changes) to be appropriate endpoints for deriving a TRV. It is also noted that, although FSANZ (2017b) did not cite the Loveless et al. (2006) study explicitly in their review, they considered increases in absolute and/or relative liver weight in rodents, in the absence of hepatocellular degeneration or necrosis, to not be an adverse effect for the purpose of identifying a NOAEL or LOAEL. Similarly, FSANZ (2017b) has not interpreted increased absolute liver weight in monkeys as an adverse effect because there was no significant effect on relative liver weight, and no histological evidence of hepatocellular hypertrophy or liver lesions.



<span id="page-76-1"></span>NJDEP (2019a), on the other hand, considered the effect appropriate for determining a guidance value and conducted BMD modelling of the serum PFOA data for the branched/linear APFO from the Loveless et al. (2006) study to derive a PFOA serum  $BMDL<sub>10</sub>$  in mice for increased relative liver weight of 4,350 ng/mL. They applied an uncertainty factor of 300 (3x for interspecies extrapolation of toxicodynamic differences, 10x for human variability, 10x for database uncertainties for potential adverse effects on mammary gland development occurring at lower doses than increased relative liver weight) to the POD to derive a target human serum level of 14.5 ng/mL. It is noted the latter database uncertainty factor was not applied in the FSANZ (2017b) derivation of a TRV and therefore would be unlikely applied if adapting the value to the Australian context. ATSDR (2021a) also noted that the mammary gland effect did not result in an adverse effect on lactational support at maternal doses of PFOA as high as 1 mg/kg/day, based on normal growth and survival in F2 pups. Given that milk production was adequate to support growth, ATSDR (2021a) considered the biological significance of the delayed development of the mammary gland observed at very low doses is uncertain.

NJDEP (2019a) then converted the target human serum level to a dose by applying a clearance factor (1.4 x 10<sup>-4</sup> L/kg/day) sourced from US EPA (2016a, as cited in NJDEP 2019a). This resulted in a TRV of 2 ng/kg/day (i.e. 14.5 ng/mL x 1.4 x 10<sup>-4</sup> L/kg/day x 10<sup>3</sup> mL/L). It is noted this TRV would likely be 10-fold higher (i.e. 20 ng/kg/day) in the Australian context if the additional database uncertainty factor was not applied (see also **Section [9.3](#page-80-0)**). In line with the conclusions in the FSANZ (2017b) review, there is uncertainty with respect to the human relevance of the liver effects observed in the Loveless et al. (2006) study due to the dearth of mode of action information for these effects and suggested human refractoriness for some of these effects. Thus, the candidate guideline value resulting from adaptation of the NJDEP (2019a) guidance value is considered to be of low confidence (see **Section [9.3](#page-80-0)**).

#### <span id="page-76-0"></span>**9.2.4 Koskela et al. (2016) – used by ATSDR (2021a)**

<span id="page-76-2"></span>Koskela et al. (2016)<sup>[18](#page-76-1)</sup> exposed pregnant C57BL/6/Bk1 mice orally in the diet to 0 (n=10) or 0.3 (n=6) mg PFOA/kg/day (96% purity) throughout pregnancy from GD1 to presumably 21, and female offspring (groups of five) were studied at age 13 or 17 months. Body weight was measured as well as morphometrical and biomechanical properties of femurs and tibias with micro-computed tomography and 3-point bending, and bone PFOA concentrations were determined by mass spectrometry. The effects of PFOA on bone cell differentiation were studied in osteoclasts from C57BL/6/Bk1 mice and in the MC3T3 pre-osteoblast cell line.

Litter mates of the offspring in the Koskela et al. (2016) study were examined for neurobehavioral effects in a study conducted by Onishchenko et al. (2011). As reported in the latter study, there were no differences in dam weight gain, litter size or sex ratio or pup body weight or brain weight at birth in the treated group compared to controls. Offspring body weight was significantly higher in comparison with controls at 13 and 17 months of age (9.9 and 7.8%, respectively), which Koskela et al. (2016) speculate may be due to an increased amount of adipose tissue.

In 17-month-old offspring, there was a 6.8% increase in periosteal area of the femoral cortical bone (p<0.05) and increases in the peri- and endosteal perimeters (3.2%, p<0.05 and 5.2%, p<0.01, respectively) and the marrow area (10.0%) (p<0.05); an increase in medullary area was also observed. There were no differences in femoral cortical bone area or femoral mineral density. In the tibia, the total area inside the periosteal envelope and the periosteal perimeter were increased (4.9 and 3.5%, respectively) (p<0.05). Although the

[<sup>18</sup>](#page-76-2) The Onishchenko et al. (2011) study (discussed in **Section 9.2.5**) and Koskela et al. (2016) study are reports of different endpoints examined in the same study.



study authors noted in the text that tibial medullary areas were "*essentially the same between groups*," data in Figure 2 of the paper show a statistically significant increase at 17 months (but absolute values were indeed similar). Significant decreases in tibial mineral density were observed at 13 and 17 months. There were no significant differences in the tibial medullary area or the endosteal perimeter. There were no significant effects on any other measured biochemical parameter in the femur or tibia (stiffness, maximum energy, absorption).

According to ATSDR (2021a), the "*Koskela et al. (2016) study has a number of strengths including examination of several measures of bone status tested at different ages, measurement of bone PFOA levels, and tests to evaluate potential mechanisms of action. To evaluate whether developmental exposure resulted in bone damage in mature animals, the study evaluated bone morphology (periosteal, cortical, and medullary areas and bone mineral density) and bone biomechanical properties (stiffness, maximum force, and maximum energy); all tests were conducted on femur and tibia bone. Measurement at two ages (13 and 17 months) allowed for an evaluation of whether the effect of PFOA on bone changed as the animals aged. The companion in vitro study of osteoclasts and osteoblasts provided mechanistic support for the in vivo findings. Additionally, the in vitro study evaluated four PFOA concentrations and found concentration-related differences."*

*There are several study limitations that affect the interpretation of the study results; these include the small number of animals tested, use of only one PFOA dose level, inadequate reporting of dietary PFOA levels, and lack of measured serum PFOA levels. Tests of potential alterations in bone mineral density and bone biomechanical properties were only evaluated in 5–6 female offspring per group; however, support for the finding comes from the consistency of the findings at 13 and 17 months of age. The use of only one PFOA dose level does not allow for the establishment of dose-response relationships. This study limitation is mitigated by the extensive intermediate-duration oral exposure database, which allows for an overall assessment of dose-response. The dams were exposed to PFOA dissolved in alcohol and sprayed onto the food pellets. The study did not measure the amount of residual alcohol or the actual amount of PFOA on the food pellets. Koskela et al. (2016) measured PFOA levels in the tibias and femurs but did not measure serum PFOA levels. ATSDR estimated the TWA serum PFOA concentrations using the Wambaugh et al. (2013) model. The lack of measured serum PFOA levels did not allow for validation of whether the model accurately predicted serum levels; the model was validated using data from other intermediate-duration PFOA studies in rats and mice* (ATSDR 2021a)".

The ATSDR (2021a) estimated mouse serum PFOA concentration at the administered dose of 0.3 mg/kg/day was 8,290 ng/mL. This serum concentration was converted by ATSDR (2021a) to a HED POD of 0.000821 mg/kg/day  $[(Css x Ke x Vd) \div AF = (8.29 mg/L) x$ 0.693/1,400 d x 0.2 L/kg  $\div$  1] and an uncertainty factor of 300 was applied (10x for use of a LOAEL, 3x for interspecies extrapolation of toxicodynamic differences, 10x for human variability).

Despite the limitations outlined by ATSDR (2021a) for the Koskela et al. (2016) study, the outcome does appear to be compelling and, if relevant to humans, could potentially increase the risk of bone fractures later in life. This study was included in the candidate guidance/guideline values summary in **Section [9.3](#page-80-0)**. However, due to the small animal numbers in the study (n=6 in treated group), the fact there was only a single treatment group, the study not following standardised testing guidelines, and the uncertainty with respect to the clinical relevance of the findings, the confidence in the resulting adapted guideline value is considered to be very low.

#### <span id="page-78-0"></span>**9.2.5 Onishchenko et al. (2011) – used by MPART (2019a)**

As described in the previous section (Koskela et al. 2016 reporting the same study), Onishchenko et al. (2011) exposed pregnant C57BL/6/Bk1 mice (n=6/group, n=10 for controls) to PFOA (96% purity) or PFOS (as heptadecafluorooctanesulfonic acid potassium salt, purity ≥ 98%) at 0 or 0.3 mg/kg/day via the diet (dissolved in ethanol and applied to food, then evaporated for 2 hours) from GD1 throughout pregnancy (presumed GD21). Tissue samples (whole brain and liver) were collected from pups at birth and concentrations of PFOS and PFOA analysed. Tests for locomotor and circadian activity were performed on offspring at age 5-8 weeks. Afterward, animals were tested for emotion-related behaviour in elevated plus maze and forced swim tests. Tests for muscle strength and motor coordination were performed in animals 3- to 4-month old.

Dams exposed to PFOS or PFOA gained weight normally during pregnancy and did not differ from control females at any gestational age. Litter size and sex ratio were unaffected by treatment and there were no differences in offspring body or brain weights between groups at birth. Liver weights were normal in PFOS-exposed pups, but significantly increased in PFOA-exposed mice  $(77 \pm 2 \text{ mg vs. } 58 \pm 1 \text{ mg in control, p<0.001}).$ 

PFOS-exposed males walked significantly less than controls when exploring a new environment, while females did not differ significantly from controls. PFOA exposure did not have a significant effect on locomotor activity. PFOS-exposed males also showed decreased activity in social groups using the TraffiCage system during the first two hours. A similar trend was observed in PFOS-exposed females, but the difference during the second hour of the test did not reach statistical significance. PFOA-exposed males were more active ( $p =$ 0.013), while PFOA-exposed females showed a decreased activity ( $p = 0.036$ ) than the controls. However, these alterations were observed when animals were tested in social groups, while individual testing did not reveal any differences. After habituation to the new home cage, animal activity declined to a low, diurnal level. All groups of animals had a normal circadian pattern with higher levels of activity during the dark phase and early morning hours, followed by lower activity levels during the light phase.

In the elevated plus maze test, PFOS-exposed male mice travelled equally long distance exploring the closed arms as controls, but the exposed animals spent significantly more time being inactive than controls. The preference for exploration of open (potentially dangerous) versus closed (safe) areas did not seem to be altered in the exposed animals. PFOSexposed females as well as all PFOA-exposed groups did not differ from their respective controls in any behavioural parameter tested in the elevated plus maze.

There was no effect on immobility time in the forced swimming test. However, in the hanging wire test, PFOS-exposed male mice had significantly shorter fall latency than controls (p =0.04) but females and PFOA-exposed mice were unaffected.

Overall, the behavioural changes found in this study were of a small magnitude and study groups were also relatively small. Serum PFOS and PFOA concentrations were not measured in this study, but ATSDR (2021a) estimated the mouse serum PFOA concentration at the administered dose of 0.3 mg/kg/day was 8,290 ng/mL. The jurisdiction did not use the results of the Onishchenko et al. (2011) study for derivation of a TRV for PFOA, since circadian activity was assessed using a TrafficCage system in which all animals in the group were placed in a single cage and activity was measured; thus, activity was only measured on a group basis and it is possible that one animal could skew the results. It is noted ATSDR (2021a) did not calculate the serum PFOS concentration in this study.

It is noted another study with a similar study design but including more than one dose (0, 0.1, 0.3, 1.0 mg/kg/day) via gavage found no changes to motor-related behaviours at PFOA doses below 1 mg/kg/day (Goulding et al. 2016).



<span id="page-79-5"></span><span id="page-79-3"></span><span id="page-79-2"></span><span id="page-79-1"></span>MPART (2019a) used both the Onishchenko et al. (2011) and Koskela et al. (2016) studies (which are reports of different endpoints examined in the same study) and considered the dose of 0.3 mg/kg/day as a LOAEL. They used the ATSDR (2021a) estimated serum concentration of 8,290 ng/mL to calculate a HED LOAEL of 0.001163 mg/kg/day [TWA serum x ke x Vd = 8.29 mg/L x 8.2 x 10<sup>-4</sup> x 0.17 L/kg]. It is noted that the parameters used to convert the serum concentration to a HED differ from those used by ATSDR (2021a).[19](#page-79-1) Using the parameters from ATSDR (2021a) results in a slightly different HED LOAEL of 0.000821 mg/kg/day. MPART (2019a) then applied the same rounded uncertainty factor of 300 (3 for use of a LOAEL since a NOAEL for immune effects was similar to the selected LOAEL and the selected LOAEL represented less severe effects, 10x for human variability, 3x for interspecies differences in toxicodynamics, 3x for database deficiencies as the mammary gland effects were considered to signal a concern for other low dose endocrine effects) to the HED LOAEL to derive a TRV of 3.9 ng/kg/day.

The study was included in the candidate guidance/guideline values summary in **Section [9.3](#page-80-0)**. However, due to the marked limitations with the study identified by ATSDR (2021a), the fact it was not conducted in accordance with standardised testing guidelines, and the apparent small absolute differences between the treated and control groups, the confidence in the resulting adapted guideline value is considered to be very low.

#### <span id="page-79-0"></span>**9.2.6 Lau et al. (2006) – used by NHMRC and NRMMC (2011), FSANZ (2017b), DOH (2017)**

Lau et al. (2006) is a developmental toxicity study conducted with PFOA (ammonium salt, >98% pure) in which timed-pregnant CD-1 mice were administered 0, 1, 3, 5, 10, 20, or 40 mg PFOA/kg bw/day by oral gavage from gestational day (GD) 1 to 17 inclusive. Some mice were sacrificed on GD18 for teratological evaluation, while others were dosed on GD18 and allowed to proceed to spontaneous parturition. In the control group, 45 mice were terminated pregnant and 23 proceeded to spontaneous parturition, whereas for the treated groups, the corresponding numbers were 17/8, 17/8, 27/19, 26/21, 42/7 and 40/0, respectively.

All dams in the 40 mg/kg/day group resorbed their litters. Weight gain in dams that carried pregnancy to term was decreased in the 20 mg/kg/day group. Increased liver weight was observed in dams sacrificed at GD18 at all doses. Percentage of live foetuses at birth was lower only in the 20 mg/kg/day group, and foetal weight was also decreased. No significant increases in malformation were noted in any treatment group. Growth was delayed in all PFOA-treated litters except the 1 mg/kg/day group. Ossification (i.e. number of sites) of the forelimb proximal phalanges was significantly decreased at all doses except 5 mg/kg/day.

<span id="page-79-4"></span>Reduced ossification of the calvaria and enlarged fontanel was observed at 1, 3, and 20 mg/kg/day and at ≥ 10 mg/kg/day in the supraoccipital bone. Postnatal survival was also significantly decreased at ≥ 5 mg/kg/day. According to the study authors, accelerated sexual maturation was observed in male offspring (i.e. time to preputial separation was decreased in male pups), but not in females, at all doses of PFOA. However, FSANZ (2017b) noted in their assessment of the study that the data presented in the paper do not support this conclusion.[20](#page-79-3)

<sup>&</sup>lt;sup>[20](#page-79-4)</sup> Age at preputial separation was similar in the high dose group (31.7  $\pm$  1.1 days) to that in controls (30.5  $\pm$  0.2 days), therefore there was no clear dose response for this effect.



<sup>&</sup>lt;sup>[19](#page-79-2)</sup> MPART (2019a) considered the PFOA serum half-life of 840 days (2.3 years) more relevant for exposure to the general population than the ATSDR (2021a) assumed half-life of 1,400 days.

<span id="page-80-1"></span>Average serum PFOA concentrations in pregnant mice at term were 21.9, 40.5, 71.9, 116, 181, and 271 µg/mL in the 1, 3, 5, 10, 20, and 40 mg/kg/day groups, respectively (FSANZ 2017b).

The maternal NOAEL was 10 mg/kg/day (i.e. 116 µg/mL), based on decreased body weight gain at  $\geq$  20 mg/kg/day (FSANZ 2017b). The NOAEL for foetal toxicity was 1 mg/kg/day (i.e. maternal serum of 21.9 µg/mL), based on decreased body weight gain at doses of ≥ 3 mg/kg/day (i.e. maternal serum of 40.5 µg/mL) (FSANZ 2017b). Lau et al. (2006) derived lower benchmark doses for a 5% effect (BMDL<sub>5</sub>s) for various effects observed in the study. The BMDL<sub>5</sub> for decreased pup weight at weaning was 0.86 mg/kg/day (serum BMDL $_5$ ) not reported), which is similar to the NOAEL nominated by FSANZ (2017b). The lowest BMDL<sup>5</sup> derived by Lau et al. (2006) was 0.17 mg/kg/day for increased maternal liver weight at term; however as noted previously, these effects, in the absence of concomitant histopathological findings, are unlikely to be relevant to humans.

FSANZ (2017b) considered the Lau et al. (2006) study suitable for derivation of a healthbased guidance value. FSANZ (2017b) used pharmacokinetic modelling to predict average serum concentrations from predicted areas-under-the-curve over the duration of dosing using parameters also used by the US EPA. The average PFOA serum concentration from the modelling at the NOAEL of 1 mg/kg/day was determined to be 35.1 µg/mL; this was converted to a human equivalent dose (HED) of 0.0049 mg/kg/day using a clearance factor of 0.00014 L/kg/day (the same factor also used by several other jurisdictions). FSANZ (2017b) then applied an uncertainty factor of 30 (3x for interspecies differences in toxicodynamics, 10x for human variability) to derive a tolerable daily intake of 160 ng/kg/day.

<span id="page-80-2"></span>The study and the resulting FSANZ (2017b) guidance value were included in the guidance / guideline values summary in **Section [9.3](#page-80-0)**. The Lau et al. (2006) study appears to have been conducted using a protocol similar to OECD TG 414 (prenatal developmental toxicity study) and examined a large number of standard endpoints<sup>[21](#page-80-1)</sup> in a sufficiently large number of treatment groups and treated animals. Thus, the confidence in the resulting guideline value is considered to be high.

### <span id="page-80-0"></span>**9.3 Candidate guidance/guideline values for PFOA**

As indicated in preceding sections, a number of additional studies (summarised in **Sections [9.2.1](#page-72-0)** to **[9.2.5](#page-78-0)**) that had not been previously explicitly considered / evaluated in the FSANZ (2017b) review of PFOA were used by various jurisdictions as critical studies for derivation of PFOA guidance values. Of those studies, all except the cross-sectional one by Gallo et al. (2012) were considered potentially suitable for adoption/adaptation for candidate DWG derivation in the Australian context.

The potentially suitable studies were used by four jurisdictions (Loveless et al. 2006 by NJDEP 2019a; Koskela et al. 2016 by ATSDR 2021a; Onishchenko et al. 2011 and Koskela et al. 2016 by MPART 2019a; and Li et al. 2017 by OEHHA 2019a) to derive a guidance value for PFOA, two of which (NJDEP 2019a and ATSDR 2021a) also met a high proportion of technical/administrative criteria for potential adoption/adaptation into the Guidelines (**Section [9.1](#page-70-0)**). However, it is noted that, due to various considerations, the confidence in the resulting adapted candidate guideline values ranges from very low to low.

The jurisdictions have all chosen different endpoints for derivation of guidance values, at times have used slightly different toxicokinetic adjustment factors for converting an animal

<sup>&</sup>lt;sup>[21](#page-80-2)</sup> Endocrine disruptor relevant parameters (i.e. anogenital distance in foetuses and thyroid hormones in dams) were only added to the OECD TG in 2018. These endpoints were not included in the Lau et al. (2006) study, since the OECD TG update superseded the conduct and publication of the Lau et al. (2006) study.



serum concentration to a human dose, and the choices of uncertainty factors also differ between jurisdictions (see **[Table 9-1](#page-82-0)**).

With respect to the relative source contribution (RSC) factor, the current factor employed in derivation of the DWGs for PFOS, PFHxS and PFOA in the Guidelines is 0.1 (i.e. 10%) which is also the default factor for the Australian context. It is noted all jurisdictions which have derived DWGs in the literature consulted applied an RSC of 0.2 (i.e. 20%) (e.g. OEHHA 2019a, 2023a; US EPA 2021a, 2022d) but do not provide the rationale for this. Thus, the default factor of 0.1 has been retained in calculating the potential resulting DWGs for PFOA using the guidance values in **[Table 9-1](#page-82-0)**, noting that it yields a lower guideline value than use of an RSC of 0.2.

Also presented in **[Table 9-1](#page-82-0)** is the derivation of the current Australian DWG for PFOA of 560 ng/L. The underpinning study on which the existing Australian PFOA guideline value is based (Lau et al. 2006) was evaluated to have high confidence in **Section [9.2.6](#page-79-0)**.

#### **Table 9-1 Potential drinking water guideline values (ng/L) resulting from adaptation of PFOA guidance values from different jurisdictions (1)**

<span id="page-82-0"></span>



#### National Health and Medical Research Council Evidence Evaluations for Australian Drinking Water Guidelines Chemical Fact Sheets – PFOS, PFHxS, PFOA, PFBS, and GenX Chemicals 17 October 2024 SLR Project No.: 640.V30693.20000



DWG = Drinking Water Guideline; BMDL = Lower Benchmark Dose; HED = Human Equivalent Dose; GD =Gestation Day. UF<sub>A</sub> = Uncertainty factor for extrapolation from animals to humans; UF<sub>H</sub> = Uncertainty factor for human variability; UF<sub>LOAEL</sub> = Uncertainty factor for use of a LOAEL rather than a NOAEL; UF<sub>composite</sub> = Composite (i.e. total) uncertainty factor; UF<sub>database</sub> = Uncertainty factor to account for the limited database of toxicological studies.  $\downarrow$  = Decreased. ↑ = Increased. APFO = ammonium perfluorooctanoate.

(1) As discussed in **Section [6.2](#page-49-0)** for PFOS, there are various reasons why the epidemiological information for associations of PFAS serum concentrations with decreased antibody titre for specific vaccines (i.e. Abraham et al. 2020, Budtz-Jørgensen and Grandjean 2018) is not considered suitable in the Australian context for derivation of guidance values for PFAS. Similarly, the cross-sectional study by Gallo et al. (2012) for increased ALT associated with increased PFOA concentrations in serum was not considered suitable for adoption/adaptation in the Australian context for PFOA health-based guidance value development (see **Section [9.2.2](#page-73-6)**). For this reason, these studies have not been included in this table.

(2) The additional uncertainty factor of 3x (for MPART 2019a) or 10x (for NJDEP 2019a) was applied for potential adverse effects on mammary gland development occurring at lower doses than the endpoint selected. As discussed in **Section [9.2.3](#page-75-0)**, this additional database uncertainty factor is unlikely to be required. The values that would result from not applying this uncertainty factor are provided in brackets.

(3) Adaptation of guidance value has been undertaken using the default assumptions for derivation of DWGs in Australia using the following equation as outlined in NHMRC (2021):

DWG (ng/L) = [Guidance value (ng/kg bw/day) x 70kg (adult) x 0.1 for adult]  $\div$  2 L/day for adult



(4) Since a NOAEL for immune effects was similar to the selected LOAEL and the selected LOAEL represented less severe effects, MPART (2019a) used a reduced uncertainty factor of 3x for use of a LOAEL.

(5) As discussed in **Section [9.2.1](#page-72-1)**, it is arguable whether the effects observed at the lowest dose in this study (0.05 mg/kg/day) in female mice can be considered adverse. If the lowest dose in the study (0.05 mg/kg/day) is considered to be a NOAEL instead of a LOAEL, the alternative values that would result are provided in brackets. In addition, FSANZ (2017b) indicates that humans may be refractory to the liver effects observed in rodents as a result of PFOA exposure, thus there is low confidence in the relevance of this candidate guideline value.

(6) As discussed in **Section [9.2.1](#page-72-1)**, the use of the additional database uncertainty factor is unlikely to be required. The values that would result from not applying this uncertainty factor are provided in brackets.

(7) The Lau et al. (2006) study appears to have been conducted using a protocol similar to OECD TG 414 (prenatal developmental toxicity study) and examined a large number of standard endpoints in a sufficiently large number of treatment groups and treated animals (see **Section [9.2.6](#page-79-5)**). Thus, the confidence in the resulting guideline value is considered to be high.

(8) Considered to be of low confidence, since increases in absolute and/or relative liver weight in rodents, in the absence of hepatocellular degeneration or necrosis, was not considered by FSANZ (2017b) to be an adverse effect for the purpose of identifying a NOAEL or LOAEL. Humans may also be more refractory to these effects than rodents (**Section 9.2.3**).

(9) Due to the small animal numbers in the Koskela et al. (2016) study (n=6 in treated group), the fact there was only a single treatment group, and the uncertainty with respect to the clinical relevance of the findings, the confidence in the resulting adapted guideline value is considered to be very low (**Section 9.2.4**).

(10) Due to the marked limitations with the Onishchenko et al. (2011) study identified by ATSDR (2021a), the fact it was not conducted in accordance with standardised testing guidelines, and the apparent small absolute differences between the treated and control groups, the confidence in the resulting adapted guideline value is considered to be very low (**Section 9.2.5**).

(11) An international collaboration of scientists (Burgoon et al. 2023) recently derived a guidance value of 70 ng/kg/day for PFOA using the same study by Lau et al. (2006). The group used a NOAEL of 23 µg/mL (i.e. 23 mg/L) and applied uncertainty factors of 2.5 for potential toxicodynamic differences between mice and humans, 3 for toxicodynamic differences between humans, and 8.4 for toxicokinetic differences between humans [23 mg/L  $\div$  63 = 0.3 mg/L]. This was converted to a guidance value by multiplying the guidance serum concentration by the geometric mean for clearance of PFOA in humans from a study by Zhang et al. (2013) assuming steady state [0.3 mg/L x 0.00023 L/day/kg = 0.00007 mg/kg/day or 70 ng/kg/day].

<span id="page-85-2"></span><span id="page-85-0"></span>The candidate PFOA DWGs derived by adapting existing guidance values for this PFAS range from 1.6 to 71 ng/L depending on the endpoint selected and uncertainty factors used, with the existing DWG at 560 ng/L. However, when excluding the values from the candidate DWGs likely not applicable to the Australian context due to differences in application of uncertainty factors or differences in endpoint selection (see **[Table 9-1](#page-82-0)**), the range is 9.6 to 71 ng/L. These values all incorporate at least an uncertainty factor of 30x in TRV development, an endpoint which is the equivalent of a dose resulting in no adverse effects, as well as a relative source contribution of 10% of the TRV to drinking water. However, it is noted that, due to various reasons outlined in **Sections [9.2.1](#page-72-0)** to **[9.2.5](#page-78-0)**, the confidence in the candidate guideline values is considered very low to low, whereas the confidence in the existing Australian guideline value is considered to be high (**Section [9.2.6](#page-79-0)**) .

It is also noted that a recently published paper by Burgoon et al. (2023) which became available at the time of writing this report describes the outcome of an international collaboration of three teams consisting of a total of 24 scientists from eight countries tasked with reviewing relevant information and independently developing ranges for estimated PFOA safe doses (i.e. guidance values). All three teams determined that the available epidemiological information could not form a reliable basis for a PFOA safe dose assessment in the absence of mechanistic data that are relevant for humans at serum concentrations seen in the general population. This conclusion is in line with the conclusions made in the current report with respect to the available epidemiological data. The international collaboration estimated PFOA guidance values ranging from 10 to 70 ng/kg/day based instead on dose-response data from five studies of PFOA-exposed laboratory animals (including the study underpinning the existing Australian guideline value, i.e. Lau et al. 2006). The collaboration considered all of these values to be protective of human health (Burgoon et al. 2023). This range of guidance values is not dissimilar from the range of PFOA guidance values shown in **[Table 9-1](#page-82-0)** adapted for the Australian context from international jurisdictions (i.e. 2.7 to 20 ng/kg/day), with the top end of the range given by Burgoon et al. (2023) (i.e. 70 ng/kg/day) being approximately two times lower than the guidance value derived by FSANZ (2017b) (i.e. 160 ng/kg/day). The difference in the latter two values is due to:

- <span id="page-85-1"></span>i) selection by Burgoon et al. (2023) of a slightly lower serum NOAEL (23 mg/L)<sup>[22](#page-85-0)</sup> than FSANZ (2017b) (35.1 mg/L) from the Lau et al. (2006) study;
- <span id="page-85-3"></span>ii) use of a slightly larger uncertainty factor by Burgoon et al. (20[23](#page-85-2)) (63 vs. 30):  $23$ and

1) 2.5x for interspecies toxicodynamic differences (instead of 3x used by FSANZ),

<sup>&</sup>lt;sup>[22](#page-85-1)</sup> It is unclear to SLR how the serum POD corresponding to the NOAEL of 23 mg/L was derived by Burgoon et al. (2023). Data summarised by FSANZ (2017b) indicates the measured serum concentration at the NOAEL dose was 21.9 mg/L, whereas FSANZ (2017b) adjusted this serum concentration to 35.1 mg/L as this was the estimated average area-under-the-curve for the duration of dosing.

 $23$  FSANZ (2017b) used a composite uncertainty factor of 30 consisting of 3x for interspecies toxicodynamic differences and 10x for human variability. Burgoon et al. (2023) used a composite uncertainty factor of 63 composed of:

<sup>2) 25.2</sup>x for human variability (instead of the default factor of 10x used by FSANZ) consisting of 3x for intra-human differences in toxicodynamics and 8.4x for intra-human differences in toxicokinetics [i.e. 0.79 mL/day/kg arithmetic mean clearance of average group from a study by Zhang et al. (2013) divided by 0.094 mL/day/kg arithmetic 95% lower bound clearance for a sensitive group from the same study = 8.4].

<span id="page-86-1"></span><span id="page-86-0"></span>iii) a slightly different human clearance value (0.00023 L/kg/day vs. 0.00014  $L/kg/day$ ).<sup>[24](#page-86-0)</sup> Collectively, these differences result in approximately a factor of  $2x$ difference in the resulting guidance values.

Since the candidate guideline values for PFOA summarised in **[Table 9-1](#page-82-0)** (9.6 to 71 ng/L) are based on data from studies considered to be of very low to low confidence for guideline derivation, it is suggested the information is not of high enough quality to warrant revision of the existing Australian guideline value for PFOA (560 ng/L) at this time, which is based on data for which there is high confidence.

In Australian distributed drinking waters or raw water catchments, PFOA concentrations generally may range up to 10 ng/L in various locations (QAEHS 2018a, 2018b, Sydney Water 2023, WHO 2022, WCWA 2023). This maximum concentration is at or below the candidate DWGs of 9.6 to 71 ng/L and well below the existing Australian guideline value of 560 ng/L. Due to the uncertainty factors and small RSC incorporated into the derivation of the candidate DWGs and the existing Australian DWG, PFOA is unlikely to present a human health risk from distributed drinking water in uncontaminated regions of Australia. However, there are many sites of PFAS contamination in Australia, and, if water from these contaminated sites is used as a local source of drinking water (e.g. backyard bore in rural location where distributed water is not available), PFOA may be present at concentrations greater than the candidate DWGs and the existing Australian DWG in these cases.

### <span id="page-86-3"></span>**10.0 Discussion for GenX Chemicals**

This section provides a discussion of the strengths and limitations of the identified guidance values for GenX Chemicals for possible adoption/adaptation into the Guidelines.

#### <span id="page-86-2"></span>**10.1 Potential suitability of health-based guidance values for possible adoption/adaptation**

Candidate guidance values for GenX Chemicals described in **Section [4.1](#page-17-0)** for possible adoption/adaptation in Australia have also been evaluated using the Assessment Tool provided in Appendix D in the Technical Report and already described in **Section [6.1](#page-48-0)** for PFOS.

**[Figure 10-1](#page-87-0)** presents the percentage of criteria (combined technical and administrative criteria) met by each jurisdiction. It is evident from the figure that the higher percentage of 'must-have' and 'should-have' criteria were met by US EPA (2021e), followed by MPART (2019a).

 $24$  FSANZ (2017b) converted the serum POD obtained from the Lau et al. (2006) study to a HED by multiplying by the clearance rate for PFOA in humans (i.e. 0.00014 L/kg/day) prior to applying the composite uncertainty factor. Burgoon et al. (2023) applied the composite uncertainty factor of 63 to the serum POD [i.e. 23 mg/L  $\div$  63 = 0.3 mg/L (rounded)] and then applied a clearance rate for PFOA in humans of 0.00023 L/kg/day [the geometric mean clearance rate from Zhang et al. (2013)] to derive the guidance value of 0.00007 mg/kg/day (i.e. 70 ng/kg/day).





#### <span id="page-87-0"></span>**Figure 10-1 Overall proportion of 'must-have', 'should-have' and 'may-have' technical/administrative criteria as per the Assessment Tool met by jurisdictions who have derived candidate guidance values for GenX chemicals for possible adoption/adaptation in Australia**

#### **10.2 Critical evaluation of GenX Chemicals guidance values**

As GenX Chemicals were not part of the comprehensive review undertaken by FSANZ (2017b), all guidance values sourced in the literature search for which the derivation was described were evaluated in this section. These include the following.

- 3 ng/kg/day (US EPA 2021e, 2022c, j; also adopted by WSDH 2022, 2023a and NJDEP 2023a) (liver effects in mice; critical study: DuPont 2010 unpublished study).
- 77 ng/kg/day (MPART 2019a) (liver effects in mice; critical study: DuPont 2010 unpublished study).

Both jurisdictions have agreed that the most sensitive health endpoint is liver effects (increased absolute and relative weight and histopathologic findings, i.e. liver single cell necrosis in parental mice) from an unpublished Reproduction/ Developmental Toxicity Study in Mice [conducted according to OECD TG 421; modified according to the Consent Order, DuPont-18405-1037 (2010)]. As the original study is not available to SLR, descriptions in the next section rely on the descriptions provided in the reviews by MPART (2019a) and US EPA (2021e).

#### <span id="page-87-1"></span>**10.2.1 DuPont (2010) – used by MPART (2019a) and US EPA (2021e)**

DuPont (2010) conducted a combined oral gavage reproductive/developmental toxicity study in mice with HFPO dimer acid ammonium salt (GenX), administering the chemical (purity 84%) to Crl:CD1(ICR) mice (25/sex/group) in deionised water at doses of 0, 0.1, 0.5, or 5 mg/kg/day. Parental males were dosed for 70 days prior to mating and throughout mating through one day prior to scheduled termination, for a total of 84-85 doses. Parental females were dosed for two weeks prior to pairing and through lactation day (LD) 20 for a total of 53- 65 doses. F1 females (offspring) were dosed daily beginning on PND21 through PND40.

US EPA evaluated the methods and data submitted as part of the DuPont (2010) unpublished study and deemed the study acceptable; they also requested an independent review of the study by the National Toxicology Program. The study was conducted according to OECD Test Guideline 421 and followed Good Laboratory Practices (GLP). This study was accompanied by additional testing also considered by the US EPA; the additional testing included repeated dose metabolism and pharmacokinetic studies in mice and rats, 90-day oral gavage toxicity study in mice and rats, and a combined chronic toxicity / carcinogenicity study in rats.

In GenX exposed males, the following effects were observed.

- ≥ 0.5 mg/kg/day: Increased absolute and relative liver weight and histopathological findings (increases in hepatocellular hypertrophy, single-cell necrosis, mitotic figures and lipofuscin pigment). Mild increases in tubular cell hypertrophy in kidneys of males.
- 5 mg/kg/day: F1 pups exhibited lower mean body weights at PND 4, 7, 14, 21, 28, 35, and 40. Delay in balanopreputial separation was also observed but was considered by US EPA to be of equivocal biological significance. Final body weight was significantly increased from controls by 9%.

In females, the following effects were observed.

- ≥ 0.5 mg/kg/day: Increased absolute and relative liver weight and histopathological findings (increases in hepatocellular hypertrophy, single-cell necrosis, mitotic figures and lipofuscin pigment).
- 5 mg/kg/day: F1 pups exhibited lower mean body weights at PND 4, 7, 14, 21, and 28. Delay in vaginal patency was also observed but was considered by US EPA to be of equivocal biological significance. Final body weight was significantly increased from controls by 14%. Increased relative kidney weight (by 6.5%) compared to controls in parental females.

Three males (one in each dose group) and six females (one in control, three in low dose, one each in mid- and high- dose groups) did not survive until scheduled sacrifice; the cause of death was undetermined in all cases except the male in the mid-dose group, which appeared to have ulcerative dermatitis. Due to the lack of dose response, the study authors concluded that these deaths were not treatment related.

No treatment-related effects were identified for reproductive parameters (mating, fertility and copulation indices; mean days between pairing and coitus). No treatment-related effects were observed for mean gestation length, mean numbers of implantation sites, mean numbers of pups born, live litter size, percentage of males at birth, postnatal survival, or general condition of pups. The NOAEL was 0.1 mg/kg/day. No plasma/serum concentration measurements were reported in the study descriptions by MPART (2019a) and US EPA (2021e).

From the study descriptions provided in MPART (2019a) and US EPA (2021e), and the independent review of the study findings by NTP, the unpublished DuPont (2010) study was conducted in accordance with relevant standardised testing guidelines and evaluated a range of endpoints. Therefore, it is concluded to be appropriate information to potentially adopt/adapt for derivation of candidate guidance/guideline values for GenX Chemicals. The candidate guidance/guideline values are summarised in **Section [10.3](#page-88-0)**.

#### <span id="page-88-0"></span>**10.3 Candidate guidance/guideline values for GenX Chemicals**

As indicated in **Section [10.2.1](#page-87-1)**, the DuPont (2010) study likely represents suitable information for potential guidance value derivation for GenX Chemicals. The study was used



<span id="page-89-1"></span>by two jurisdictions (MPART 2019a; US EPA 2021e; the latter also adopted by WSDH 2022, 2023a and NJDEP 2023a 2023a) to derive a guidance value for GenX Chemicals, of which US EPA (2021e) met a higher proportion of technical/administrative criteria for potential adoption/adaptation into the Guidelines (**Section [10.1](#page-86-2)**).

The two jurisdictions who derived a guidance value for GenX Chemicals using the DuPont (2010) study used similar PODs; MPART (2019a) used a  $BMDL_{10}$  of 0.15 mg/kg/day for liver single cell necrosis, whereas US EPA (2021e) used a BMD $L_{10}$  of 0.09 mg/kg/day for the constellation of liver lesions in parental females. Both jurisdictions used an allometric scaling approach to translate the POD to a HED POD<sup>[25](#page-89-1)</sup> by applying a factor of 0.15 to the POD. This gave HED PODs of 0.01 mg/kg/day (US EPA 2021e) or 0.0225 mg/kg/day (MPART 2019a).

<span id="page-89-2"></span>The jurisdictions then applied different uncertainty factors (300 or 3,000) to their HED POD (see **[Table 10-1](#page-89-0)**). The difference is due to application of an additional uncertainty factor of 10 by US EPA (2021e) for database uncertainties. However, as discussed for PFHxS in **Section [7.3](#page-58-0)**, it is not considered warranted to apply full uncertainty factors of 10x each for both the use of a subchronic study and database uncertainties.

With respect to the relative source contribution (RSC) factor, the current factor employed in derivation of the DWGs for PFOS, PFHxS and PFOA in the Guidelines is 0.1 (i.e. 10%) which is also the default factor for the Australian context. It is noted that both jurisdictions which have derived DWGs in the literature consulted applied an RSC of 0.2 (i.e. 20%) (e.g. MPART 2019a, US EPA 2022j) but do not provide the rationale for this. Thus, the default factor of 0.1 has been retained in calculating the potential resulting DWGs for GenX Chemicals using these guidance values in **[Table 10-1](#page-89-0)**, noting that it yields a lower guideline value than use of an RSC of 0.2.

<span id="page-89-0"></span>

<b>Parameter</b>	<b>MPART 2019a</b>	US EPA 2021e, 2022c, j; also adopted by WSDH 2022, 2023a and NJDEP 2023a
Critical study	DuPont 2010	
Study population	Mice	
Form of GenX studied	HFPO dimer acid ammonium salt	
Exposure route	Oral (gavage)	
Study timeframe	Combined reproductive/developmental toxicity (Parental males = 70 days prior to mating and throughout mating through one day prior to scheduled termination for a total of 84-85 doses. Parental females = two weeks prior to pairing and through LD 20 for a total of 53-65 doses. F1 females (offspring) = daily beginning on PND21 through PND40).	
<b>Critical Effect</b>	Liver single cell necrosis in parental males	Constellation of liver lesions in parental females
Point of Departure (mg/kg/day)	$BMDL_{10} = 0.15$	$BMDL_{10} = 0.09$

<sup>&</sup>lt;sup>[25](#page-89-2)</sup> The approach involves BW<sup>3/4</sup> scaling, i.e. (body weight in animal<sup>{4}</sup> ÷ body weight in human<sup>{2}</sup>) = [(0.0372 kg in male mouse)<sup>1/4</sup>  $\div$  (80 kg)<sup>1/4</sup>] = 0.15. If the convention of 70 kg used in the Guidelines were used in this equation, the factor of 0.15 would remain unchanged, so this has no influence on the POD.



DWG = Drinking Water Guideline; BMDL = Lower Benchmark Dose; HED = Human Equivalent Dose; LD = Lactation Day; PND = Postnatal Day; UF<sub>A</sub> = Uncertainty factor for extrapolation from animals to humans; UF<sub>H</sub> = Uncertainty factor for human variability;  $UF_{timeframe} = Uncertainty$  factor for use of a short-term study;  $UF_{composite}$  $=$  Composite (i.e. total) uncertainty factor; UF $_{\text{database}}$  = Uncertainty factor to account for the limited database of toxicological studies (e.g. no two-generation or immunotoxicity studies).

(1) Adaptation of guidance value has been undertaken using the default assumptions for derivation of DWGs in Australia using the following equation as outlined in NHMRC (2021):

DWG (ng/L) = [Guidance value (ng/kg bw/day) x 70kg (adult) x 0.1 for adult]  $\div$  2 L/day for adult (2) The approach involves BW $^{3/4}$  scaling, i.e. (body weight in animal<sup> $\frac{1}{4}$ </sup> ÷ body weight in human<sup> $\frac{1}{4}$ </sup>) = [(0.0372 kg in male mouse)<sup>1/4</sup> ÷ (80 kg)<sup>1/4</sup>] = 0.15. If the convention of 70 kg used in the Guidelines were used in this

equation, the factor of 0.15 would remain unchanged, so this has no influence on the POD.

The candidate GenX Chemicals DWGs derived by adapting existing guidance values for this PFAS are 263 ng/L using the uncertainty factors used by MPART (2019a) or 12 ng/L using the additional uncertainty factor employed by US EPA (2021e). As discussed in the text preceding the table, the main difference between the two values is the application of higher uncertainty factors (10 each for timeframe and database deficiencies by US EPA 2021e, 2022c, j; but only 3 each by MPART 2019a).

However, it is noted that there is only one toxicological study available on which to base a candidate DWG. There is also concern with respect to the reported purity (i.e. 84%) of GenX in the DuPont (2010) study. Therefore, a value of 263 ng/L could be regarded as a concentration of potential concern rather than a DWG *per se*.

Unfortunately, no information regarding GenX Chemical levels in Australian distributed drinking water was identified from the literature retrieved. Therefore, it is unknown whether GenX Chemicals are present at concentrations lower or higher than the concentration of potential concern. It is suggested additional research is needed to determine whether GenX Chemicals are found in any Australian drinking waters, which would also inform whether a health-based DWG is required.

## **11.0 Conclusions**

The targeted screening of existing health-based guidance/guideline values for the five PFAS of interest identified numerous candidate guidance/guideline values for potential adoption/adaptation.

The volume of information found and needing to be assessed was very large. Due to resource constraints and with agreement from NHMRC with advice from the Committee, critical evaluation of studies was prioritised to those studies that had not been previously reviewed and/or considered by an Australian agency for guidance/guideline value development. The latest review by an Australian jurisdiction in which guidance values were derived for three of the PFAS under consideration (PFOS+PFHxS and PFOA) was the FSANZ (2017b) document. This forms the basis of the current TRVs for PFOS/PFHxS and PFOA which have been used by NHMRC to derive the current guideline values in drinking water for these chemicals. FSANZ (2021) also published a review of immunomodulation effects, in which the jurisdiction reviewed a number of studies, findings of which are used to support discussions in this report on relevant PFAS.

A summary of the conclusions and DWG options from potential adoption/adaptation of suitable information for each of the five PFAS is provided in **[Table 11-1](#page-91-0)**. Bolded guideline values in the table below are considered to be most relevant to the Australian context in terms of selection of uncertainty factors and endpoints.



#### <span id="page-91-0"></span>**Table 11-1 Conclusions and DWG options from potential adoption/adaptation of suitable information for PFOS, PFHxS, PFBS, PFOA, and GenX Chemicals**











DWG = Drinking Water Guideline. TRV = Toxicity Reference Value. UF = Uncertainty Factor. RSC = Relative Source Contribution.

(1) Values that are **bolded** are considered to be most relevant to the Australian context in terms of selection of uncertainty factors and endpoints (see detailed discussions in **Section [6.0](#page-48-1)** to **[10.0](#page-86-3)** for further information).

(2) These are values that would result from a change to the selected uncertainty factors and/or endpoint type by a particular jurisdiction; the suggested changes are considered to be in line with the Australian context such as to provide consistency with the approach taken to uncertainty considerations by FSANZ (2017b). However, it is noted that the candidate guideline values for PFOA (9.5 to 70 ng/L) are based on data from studies considered to be of very low to low confidence for guideline derivation.

From the available information gathered on exposure to the five PFAS of interest in Australian distributed drinking waters and the information gathered to inform supporting information in the Fact Sheet, all DWG options would be readily measurable with current commercial analytical techniques. Although existing treatment technologies do not appear to be particularly effective at removing PFAS from water, DWG options are/would be achievable if uncontaminated source waters are utilised. However, the DWG options may not be achievable for local drinking water supplies in contaminated areas without addition of a PFAS-removal treatment step or use of an alternative water supply.

### **12.0 Review Team**





## **13.0 Declared Interests**



### **14.0 Acknowledgements**

The authors acknowledge NHMRC and the members of the Committee for their insightful review comments.

### **15.0 References**

Abraham, K., Mielke, H., Fromme, H., Volkel, W., Menzel, J., Peiser, M., Zepp, F., Willich, S.N. and Weikert, C., (2020). Internal exposure to perfluoroalkyl substances (PFASs) and biological marker in 101 healthy 1-year-old children: associations between levels of perfluorooctanoic acid (PFOA) and vaccine response. Archives of Toxicology, 94, 2131– 2147.

AECOM (2017). Stage 2C Environmental Investigation - Human Health Risk Assessment – 2017. Army Aviation Centre Oakey (AACO), Oakey QLD. 1 December 2017. 60533675 Revision 0 Final. AECOM Australia Pty Ltd (AECOM).

AECOM (2017b). Off-site Human Health Risk Assessment. December 2017. RAAF Base Williamtown. Stage 2B Environmental Investigation. 1 December 2017. 60527153. Revision 1 - Final. AECOM Australia Pty Ltd (AECOM).

Alaska DEC (2019a). Technical Memorandum Action Levels for PFAS in Water and Guidance on Sampling Groundwater and Drinking Water. Date: August 20, 2018, Updated: October 2, 2019. Division of spill prevention and response. Contaminated sites program and Division of environmental health Drinking water program. Alaska Department of Environmental Conservation (Alaska DEC).

ALS (2023). RE: [EXTERNAL] – RE: PFAS – RE: QUOTE AS ATTACHED – ALS website enquiry. Email from ALS Client Manager, Water on 24 August 2023. Eurofins.

ANU (2022). PFAS Health Study FAQs. Australian National University. Published 18 January 2022.

https://nceph.anu.edu.au/files/ANU%20PFAS%20Health%20Study%20Website%20FAQ\_22 0118.pdf#overlay-context=research/projects/pfas-health-study/reports

ATSDR (2018a). ATSDR's Minimal Risk Levels (MRLs) and Environmental Media Evaluation Guides (EMEGs) for Perfluoroalkyls (PFAS). November 2018. Agency for Toxic Substances and Disease Registry (ATSDR).

ATSDR (2021a). Toxicological Profile for Perfluoroalkyls. Released May 2021. Agency for Toxic Substances and Disease Registry (ATSDR).

Ballesteros V., Costa O., Iñiguez C., Fletcher T., Ballester F. and Lopez-Espinosa M. J. (2017). Exposure to perfluoroalkyl substances and thyroid function in pregnant women and children: A systematic review of epidemiologic studies. Environment International 99: 15-28.

Benskin J. P., De Silva A. O., Martin L. J., Arsenault G., McCrindle R., Riddell N., Mabury S. A. and Martin J. W. (2009). Disposition of perfluorinated acid isomers in Sprague-Dawley rats; Part 1: single dose. Environmental Toxicology and Chemistry 28(3): 542-567.

BfR (2019a). New health-based guidance values for the industrial chemicals PFOS and PFOA BfR opinion No 032/2019 of 21 August 2019. German Federal Institute for Risk Assessment. Bundesinstitut für Risikobewertung (BFR).

Bil, W., Zeilmaker, M., Fragki, S., Lijzen, J., Verbruggen, E., Bokkers, B. (2021). Risk Assessment of Per- and Polyfluoroalkyl Substance Mixtures: A Relative Potency Factor Approach. Environmental Toxicology and Chemistry, 40, 859-870. DOI: 10.1002/etc.4835 (as quoted in RIVM 2021a).

Boesen S. A. H., Long M., Wielsøe M., Mustieles V., Fernandez M. F. and Bonefeld-Jørgensen E. C. (2020). Exposure to Perflouroalkyl acids and foetal and maternal thyroid status: a review. Environmental Health 19(1): 107.

BSC (2021). Burdekin Shire Council. Drinking Water Management Plan. V3.3. 1 February 2021. Burdekin Shire Council (BSC).

Budtz-Jørgensen, E., and P. Grandjean. (2018). Application of benchmark analysis for mixed contaminant exposures: mutual adjustment of perfluoroalkylate substances associated with immunotoxicity. PloS One 13(10):e0205388.

Burgoon L. D., Clewell H. J., Cox T., Dekant W., Dell L. D., Deyo J. A., Dourson M. L., Gadagbui B. K., Goodrum P., Green L. C., Vijayavel K., Kline T. R., House-Knight T., Luster M. I., Manning T., Nathanail P., Pagone F., Richardson K., Severo-Peixe T., Sharma A., Smith J. S., Verma N. and Wright J. (2023). Range of the perfluorooctanoate (PFOA) safe dose for human health: An international collaboration. Regulatory Toxicology and Pharmacology 145: 105502.

Butenhoff, J., Costa, G., Elcombe, C., Farrar, D., Hansen, K., Iwai, H., Jung, R., Kennedy, Jr. G., Lieder, P., Olsen, G. and Thomford, P. (2002) Toxicity of Ammonium Perfluorooctanoate in Male cynomolgus Monkeys after Oral Dosing for 6 Months. Toxicol Sci 69: 244–257

Butenhoff, J.L., Chang, S-C., Ehresman, D.J., York RG (2009). Evaluation of potential reproductive and developmental toxicity of potassium perfluorohexanesulfonate in Sprague Dawley rats. Reprod Toxicol 27(3-4): 331-341.

Butenhoff, J.L., Chang, S.C., Olsen, G.W. and Thomford, P.J. (2012b). Chronic dietary toxicity and carcinogenicity study with potassium perfluorooctanesulfonate in Sprague Dawley rats. Toxicology, 293(1–3): 1–15 (as quoted in HC 2019a).

CDPH (2023a). Per- and polyfluoroalkyl Substances (PFAS). 2023. Connecticut State Department of Public Health (CDPH).

Chang, S., Butenhoff, J.L., Parker, G.A., Coder, P.S., Zitzow, J.D., Krisko, R.M., Bjork, J.A., Wallace, K.B., Seed, J.G. (2018). Reproductive and developmental toxicity of potassium perfluorohexanesulfonate in CD-1 mice. Reprod Toxicol 78: 150-168.

Coperchini F., Croce L., Ricci G., Magri F., Rotondi M., Imbriani M. and Chiovato L. (2020). Thyroid Disrupting Effects of Old and New Generation PFAS. Frontiers in Endocrinology (Lausanne) 11: 612320.

Corton J. C., Peters J. M. and Klaunig J. E. (2018). The PPARα-dependent rodent liver tumor response is not relevant to humans: addressing misconceptions. Arch Toxicol 92(1): 83-119.

DHAC (2023). Fundamentals of Immunisation, Australian Government Department of Health and Aged Care, Australian Immunisation Handbook. [Accessed 14/09/2023]. https://immunisationhandbook.health.gov.au/contents/fundamentals-ofimmunisation#vaccine-efficacy-and-vaccine-effectiveness.

DOH (2017). Health Based Guidance Values for PFAS. 2017. Department of Health (DOH), Australian Government.

Dong G.-H., Zhang Y.-H., Zheng L., Liu W., Jin Y.-H. and He Q.-C. (2009). Chronic effects of perfluorooctanesulfonate exposure on immunotoxicity in adult male C57BL/6 mice. Archives of Toxicology 83(9): 805-815.

Dong, G., Liu, M.M., Wang, D., Zheng, L., Liang, Z.F., Jin, Y.H. (2011). Sub-chronic effect of perfluorooctanesulfonate (PFOS) on the balance of type 1 and type 2 cytokine in adult C57BL6 mice. Archives of Toxicology 85: 1235-1244.

Drew, R. and Hagen, T. (2016) Immunomodulation by PFASs: A brief literature review. ToxConsult document ToxCR300816. (As quoted in FSANZ 2017b).

DuPont (2010). Oral (gavage) reproduction/developmental toxicity study in mice (OECD TG 421; modified according to the Consent Order) DuPont-18405-1037. Unpublished. As cited in MPART 2019a.



EC (2022). Final Opinion on Groundwater quality standards for proposed additional pollutants in the annexes to the Groundwater Directive (2006/118/EC) Scientific Committee on Health, Environmental and Emerging Risks SCHEER. 18 July 2022. DG Health and Food Safety. European Commission (EC).

EFSA (2020a). Risk to human health related to the presence of perfluoroalkyl substances in food. Adopted: 9 July 2020. European Food Safety Authority (EFSA).

enHealth (2016). enHealth Statement: Interim national guidance on human health reference values for per- and poly-fluoroalkyl substances for use in site investigations in Australia, Environmental Health Standing Committee (enhealth) of the Australian Health Protection Principal Committee. June 2016.

EU (2020). Directives. Directive (EU) 2020/2184 of the European Parliament and of the Council of 16 December 2020 on the Quality of Water Intended for Human Consumption (recast) (Text with EEA relevance). L 435/2, 23.12.2020. European Union (EU).

Eurofins (2023). RE: AU\_Environment\_Enquiry. Email from Eurofins Business Development Manager VIC, TAS, NT, SA on 22 August 2023. Eurofins.

Feng, X; Cao, X; Zhao, S; Wang, X; Hua, X; Chen, L; Chen, L. (2017). Exposure of pregnant mice to perfluorobutanesulfonate causes hypothyroxinemia and developmental abnormalities in female offspring. Toxicol Sci 155: 409-419.

FSANZ (2017b). Hazard assessment report – Perfluorooctane Sulfonate (PFOS), Perfluorooctanoic Acid (PFOA), Perfluorohexane Sulfonate (PFHxS). Food Standards Australia New Zealand (FSANZ).

FSANZ (2021). PFAS and Immunomodulation Review and Update, Food Standards Australia New Zealand.

Gallo, V., Leonardi, G., Genser, B., Lopez-Espinosa, M.J., Frisbee, S.J., Karlsson, L., Ducatman, A.M., Fletcher, T. (2012). Serum perfluorooctanoate (PFOA) and perfluorooctane sulfonate (PFOS) concentrations and liver function biomarkers in a population with elevated PFOA exposure. Environ Health Perspect 120(5): 655-660.

GHD (2018). RAAF Base Pearce PFAS Investigation. Human Health Risk Assessment Consolidated Report. July 2018. GHD Pty Ltd (GHD).

Goulding D. R., White S. S., McBride S. J., Fenton S. E. and Harry G. J. (2016). Gestational exposure to perfluorooctanoic acid (PFOA): alterations in motor related behaviors. NeuroToxicology 58: 110-119.

Grandjean, P., E.W. Andersen, E. Budtz-Jørgensen, F. Nielsen, K. Mølbak, P. Weihe, and C. Heilmann. (2012). Serum vaccine antibody concentrations in children exposed to perfluorinated compounds. JAMA 307:391–397 (as quoted in USEPA 2021d)

Grandjean P, Heilmann C, Weihe P, Nielsen F, Mogensen UB, Timmermann A, et al. Estimated exposures to perfluorinated compounds in infancy predict attenuated vaccine antibody concentrations at age 5-years. J Immunotoxicol. 2017; 14(1): 188–95. https://doi.org/10.1080/1547691X.2017.1360968 PMID: 28805477. (As cited in Buftz-Jørgensen and Grandjean 2018).

Hall, A.P., Elcombe, C.R., Foster, J.R., Harada, T., Kaufmann, W., Knippe, ,l.A., Küttler, K., Malarkey, D.E., Maronpot, R.R., Nishikawa, A., Nolte, T., Schulte, A., Strauss, V. and York, M.J. (2012) Liver Hypertrophy: A Review of Adaptive (Adverse and Non-adverse) Changes – Conclusions from the 3rd International ESTP Expert Workshop. Toxicol Pathol 40: 971-994.

HC (2018a). Guidelines for Canadian Drinking Water Quality Guideline Technical Document Perfluorooctane Sulfonate (PFOS). December 2018. Health Canada (HC). Government of Canada.

HC (2018b). Guidelines for Canadian Drinking Water Quality Guideline Technical Document Perfluorooctanoic Acid (PFOA). December 2018. Health Canada (HC). Government of Canada.

HC (2019a). Water talk: Summary of drinking water values for PFOS, PFOA and other PFAS. Health Canada (HC). Government of Canada.

Kirk M, Smurthwaite K, Braunig J, Trevenar S, D'Este C, Lucas R, Lal A, Korda R, Clements A, Mueller J, Armstrong B (2018). The PFAS Health Study: Systematic Literature Review. Canberra: The Australian National University.

https://nceph.anu.edu.au/files/PFAS%20Health%20Study%20Systematic%20Review\_1.pdf

Koskela, A., Finnilä, M.A., Korkalainen, M., Spulber, S., Koponen, J., Håkansson, H., Tuukkanen, J., Viluksela, M.. (2016). Effects of developmental exposure to perfluorooctanoic acid (PFOA) on long bone morphology and bone cell differentiation. Toxicol. Appl. Pharmacol. 301:14-21.

Lau, C., Thibodeaux, J.R., Hanson, R.G., Narotsky M.G., Rogers, J.M., Lindstrom, A.B. and Strynar, M.J. (2006). Effects of Perfluorooctanoic Acid Exposure during Pregnancy in the Mouse. Toxicol Sci 90 510–518

Lau C., Rumpler J., Das K. P., Wood C. R., Schmid J. E., Strynar M. J. and Wambaugh J. F. (2020). Pharmacokinetic profile of Perfluorobutane Sulfonate and activation of hepatic nuclear receptor target genes in mice. Toxicology 441: 152522.

Li K., Gao P., Xiang P., Zhang X., Cui X. and Ma L. Q. (2017). Molecular mechanisms of PFOA-induced toxicity in animals and humans: Implications for health risks. Environment International 99: 43-54.

Loveless, S.E., Finlay, C., Everds, N.E., Frame, S.R., Gillies, P.J., O'Connor, J.C., Powley, C.R., Kennedy, G.L. (2006). Comparative responses of rats and mice exposed to linear/branched, linear, or branched ammonium perfluorooctanoate (APFO). Toxicology 220: 203–217.

Luebker, D.J., Case, M.T., York, R.G., Moore, J.A., Hansen, K.J., Butenhoff, J.L. (2005a). Two-generation reproduction and cross-foster studies of perfluorooctanesulfonate (PFOS) in rats. Toxicol 215:126-148.

Luebker D. J., York R. G., Hansen K. J., Moore J. A. and Butenhoff J. L. (2005b). Neonatal mortality from in utero exposure to perfluorooctanesulfonate (PFOS) in Sprague–Dawley rats: Dose–response, and biochemical and pharamacokinetic parameters. Toxicology 215(1–2): 149-169.

Maine DHHS (2021a). PFOS, PFOA and other PFAS Questions and Answers. Updated 7/07/2021. Maine Department of Health and Human Services (Maine DHHS).

Mass DEP (2022a). Important Information. EPA's New Health Advisories for Some PFAS. August 11, 2022. Department of Environment Protection. Commonwealth of Massachusetts (Mass DEP).

Mayo Clinic (2023). Liver function tests. [Accessed 14/09/2023]. https://www.mayoclinic.org/tests-procedures/liver-function-tests/about/pac-20394595#:~:text=ALT%20.%207%20to%2055%20units,40%20to%20129%20U%2FL%20.

MDH (2020a). Toxicological Summary for: Perfluorooctane sulfonate. August 2020. Health-Based Guidance for Water. Health Risk Assessment Unit, Environmental Health Division. Minnesota Department of Health (MDH).

MDH (2020b). Toxicological Summary for: Perfluorohexane sulfonate. August 2020. Health-Based Guidance for Water. Health Risk Assessment Unit, Environmental Health Division. Minnesota Department of Health (MDH).

MDH (2022d). PFOA and Water. April 2022. Minnesota Department of Health (MDH).

MDH (2022f). Toxicological Summary for: Perfluorooctanoate. March 2022. Health-Based Guidance for Water. Health Risk Assessment Unit, Environmental Health Division. Minnesota Department of Health (MDH)

MDH (2022e). PFBS and Drinking Water. March 2022. Minnesota Department of Health (MDH).

MDH (2022g). Toxicological Summary for: Perfluorobutane sulfonate. March 2022. Health-Based Guidance for Water. Health Risk Assessment Unit, Environmental Health Division. Minnesota Department of Health (MDH).

MDH (2023a). News Release. Joint agency statement on draft federal limits on PFAS in drinking water. March 14, 2023. Minnesota Department of Health (MDH).

Moher D., Liberati A., Tetzlaff J. and Altman D. G. (2009). Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. BMJ 339: b2535.

MPART (2019a). Health-Based Drinking Water Value Recommendations for PFAS in Michigan. June 27, 2019. Michigan Science Advisory Workgroup. Michigan's PFAS Action Response Team (MPART).

NC DHHS (2017). Gen X Health Information. 2017. State of North Carolina. Department of Health and Human Services (NC DHHS).

NHMRC and NRMMC (2011). Australian Drinking Water Guidelines 6 2011; Version 3.8 updated September 2022, National Health and Medical Research Council and Natural Resource Management Ministerial Council, Commonwealth of Australia, Canberra.

NMI (2023). Perfluoroalkyl and polyfluoroalkyl substances (PFASs) in Water. In PDF file titled "NSW PFAS Capability in Water\_2023.pdf". National Measurement Institute (NMI).

NTP (2018). TOX-96: Toxicity Report Tables and Curves for Short-term Studies: Perfluorinated Compounds: Sulfonates and personal communication between MDH and NTP project manager Dr. Chad Blystone (as cited in the HRA Toxicology Review Worksheet for PFHxS, last revised 3/8/2019). National Toxicology Program (NTP).

NTP (2019). NTP Technical Report on the Toxicity Studies of Perfluoroalkyl Sulfonates (Perfluorobutane Sulfonic Acid, Perfluorohexane Sulfonate Potassium Salt, and Perfluorooctane Sulfonic Acid) Administered by Gavage to Sprague Dawley Rats P.H. Service, Editor. 2019, National Toxicology Program (NTP), U.S. Department of Health and Human Services: Research Triangle Park, NC.

NTP (2022). NTP technical report on the toxicity studies of perfluoroalkyl sulfonates (perfluorobutane sulfonic acid, perfluorohexane sulfonate potassium salt, and perfluorooctane sulfonic acid) administered by gavage to Sprague Dawley (Hsd:Sprague Dawley SD) rats (revised). Research Triangle Park, NC: National Toxicology Program. Toxicity Report 96.

NJDEP (2019a). Technical Support Document: Interim Specific Ground Water Criterion for Perfluorooctanoic Acid (PFOA, C8) (CAS #: 335-67-1; Chemical Structure: CF3(CF2)6COOH)\*. March 6, 2019. New Jersey Department of Environmental Protection (NJDEP).

NJDEP (2019b). Technical Support Document: Interim Specific Ground Water Criterion for Perfluorooctane Sulfonate (PFOS) (CAS #: 1763-23-1; Chemical Formula: C8HF17O3S). March 6, 2019. New Jersey Department of Environmental Protection (NJDEP).

NJDEP (2023a). Interim Specific Ground Water Quality Criterion (ISGWQC) for hexafluoropropylene oxide dimer acid (HFPO-DA) and its ammonium salt (GenX). May 24, 2023. Department of Environmental Protection. State of New Jersey (NJDEP).

OEHHA (2019a). Notification Level Recommendations. Perfluorooctanoic Acid and Perfluorooctane Sulfonate in Drinking Water. August 2019. Pesticide and Environmental Toxicology Branch. Office of Environmental Health Hazard Assessment (OEHHA). California Environmental Protection Agency.

OEHHA (2021d). Notification Level Recommendation. Perfluorobutane Sulfonic Acid in Drinking Water. January 2021. Pesticide and Environmental Toxicology Branch. Office of Environmental Health Hazard Assessment (OEHHA). California Environmental Protection Agency.

OEHHA (2022a). Notification Level Recommendation. Perfluorohexane Sulfonic Acid in Drinking Water. March 2022. Pesticide and Environmental Toxicology Branch. Office of Environmental Health Hazard Assessment (OEHHA). California Environmental Protection Agency.

OEHHA (2023a). Public Health Goals. Second Public Review Draft. Perfluorooctanoic Acid and Perfluorooctane Sulfonic Acid in Drinking Water. July 2023. Pesticide and Environmental Toxicology Branch. Office of Environmental Health Hazard Assessment (OEHHA). California Environmental Protection Agency.

Onishchenko, N., Fischer, C., Wan Ibrahim, W.N., Negri, S., Spulber, S., Cottica, D., Ceccatelli, S. (2011). Prenatal exposure to PFOS or PFOA alters motor function in mice in a sex-related manner. Neurotox. Res. 19(3):452-61.

Perkins, R., Butenhoff, J., Kennedy, G. and Palazzolo, M. (2004). 13-Week dietary toxicity study of ammonium perfluorooctanoate (APFO) in male rats. Drug and Chemical Toxicology 27: 361-378.

RIVM (2021a). Revised Risk Assessment of GenX And PFOA in Food. Part 1: Toxicity of GenX and PFOA and intake through contaminated Dairy products, eggs and fish. 01-09- 2021 (final version). Rijksinstituut voor Volksgezondheid en Milieu (RIVM).

QAEHS (2018a). Catchment and Drinking Water Quality Micro Pollutant Monitoring Program – Passive Sampling. Report 8 – Summer 2018. Queensland Alliance for Environmental Health Sciences (QAEHS).

QAEHS (2018b). Catchment and Drinking Water Quality Micro Pollutant Monitoring Program – Passive Sampling. Report 9 – Winter 2018. Queensland Alliance for Environmental Health Sciences (QAEHS).

SGS (2023). RE: [EXTERNAL] RE: Summary of methods for metals in drinking water. Email from SGS Key Account Manager on 22 August 2023. SGS Australia (SGS).

Steenland, K., Tinker, S., Frisbee, S., Ducatman, A., Vaccarino, V. (2009): Association of perfluorooctanoic acid and 101erfluorooctane sulfonate with serum lipids among adults living near a chemical plant. Am J Epidemiol 170(10):1268-78.

Sydney Water (2023). PFAS and Drinking Water. Sydney Water. Last accessed on 06 September 2023 at this location: https://www.sydneywater.com.au/water-theenvironment/how-we-manage-sydneys-water/safe-drinking-water/water-analysis/pfas-anddrinking-water.html.

ToxConsult (2019). Assessment of International and National Agency processes for deriving HBGVs and DWGs. Prepared for National Health and Medical Research Council. ToxConsult document: ToxCR070519-TF, dated 24<sup>th</sup> December 2019.

USEPA (2018). Human Health Toxicity Values for Perfluorobutane Sulfonic Acid (CASRN 375-73-5) and Related Compound Potassium Perfluorobutane Sulfonate (CASRN 29420-49- 3). Public Comment Draft. EPA-823-R-18-307. USEPA, Office of Research and Development, Washington, DC. Accessed online June 1, 2019. United States Environmental Protection Agency (USEPA).

USEPA (2021a). External Peer Review Draft. Proposed Approaches to the Derivation of a Draft Maximum Contaminant Level Goal for Perfluorooctanoic Acid (PFOA) (CASRN 335-67- 1) in Drinking Water. EPA Document No. 822D21001. December 2021. DRAFT. DO NOT CITE OR QUOTE. United States Environmental Protection Agency (USEPA).

USEPA (2021b). External Peer Review Draft. Proposed Approaches to the Derivation of a Draft Maximum Contaminant Level Goal for Perfluorooctane Sulfonic Acid (PFOS) (CASRN 1763-23-1) in Drinking Water. EPA Document No. 822D21002. December 2021. DRAFT. DO NOT CITE OR QUOTE. United States Environmental Protection Agency (USEPA).

USEPA (2021c). Human Health Toxicity Values for Perfluorobutane Sulfonic Acid (CASRN 375-73-5) and Related Compound Potassium Perfluorobutane Sulfonate (CASRN 29420-49- 3). EPA/600/R-20/345F. April 2021. United States Environmental Protection Agency (USEPA).

USEPA (2021e). Human Health Toxicity Values for Hexafluoropropylene Oxide (HFPO) Dimer Acid and Its Ammonium Salt (CASRN 13252-13-6 and CASRN 62037-80-3). Also Known as "GenX Chemicals". EPA-Final. EPA Document Number: 822R-21-010. October 2021. United States Environmental Protection Agency (USEPA).

USEPA (2022c). Technical Fact Sheet: Drinking Water Health Advisories for Four PFAS (PFOA, PFOS, GenX chemicals, and PFBS). EPA Document No. EPA 822-F-22-002. June 2022. United States Environmental Protection Agency (USEPA).

USEPA (2022d). Interim Drinking Water Health Advisory: Perfluorooctanoic Acid (PFOA) CASRN 335-67-1. EPA Publication # EPA/822/R-22/003. June 2022. United States Environmental Protection Agency (USEPA).

USEPA (2022e). Interim Drinking Water Health Advisory: Perfluorooctane Sulfonic Acid (PFOS) CASRN 1763-23-1. EPA Publication # EPA/822/R-22/004. June 2022. United States Environmental Protection Agency (USEPA).

USEPA (2022j). Drinking Water Health Advisory: Hexafluoropropylene Oxide (HFPO) Dimer Acid (CASRN 13252-13-6) and HFPO Dimer Acid Ammonium Salt (CASRN 62037-80-3), Also Known as "GenX Chemicals". EPA-Final. EPA Document Number: EPA/822/R-22/005. June 2022. United States Environmental Protection Agency (USEPA).

USEPA (2022k). Drinking Water Health Advisory: Perfluorobutane Sulfonic Acid (CASRN 375-73-5) and Related Compound Potassium Perfluorobutane Sulfonate (CASRN 29420-49- 3). EPA/822/R-22/006. June 2022. United States Environmental Protection Agency (USEPA).

USEPA (2023). IRIS Toxicological Review of Perfluorohexanesulfonic Acid (PFHxS, CASRN 335-46-4) and Related Salts. EPA Publication # EPA/635/R-23/148a. External Review Draft. July 2023. United States Environmental Protection Agency (USEPA).

WCWA (2019). Drinking Water Quality. Annual Report 2018-19. 2019. Water Corporation of Western Australia (WCWA).

WCWA (2020). Drinking Water Quality. Annual Report 2019-20. 2020. Water Corporation of Western Australia (WCWA).

WCWA (2021). Drinking Water Quality. Annual Report 2020-21. 2021. Water Corporation of Western Australia (WCWA).



WCWA (2023). Advice Article. PFAS & Esperance Town Water Supply Scheme. 2023. Water Corporation of Western Australia (WCWA). Last accessed online on 06 September 2023 at this location: https://www.watercorporation.com.au/Help-and-advice/Waterissues/Water-quality/Known-water-issues/PFAS-and-Esperance-Town-Water-Supply-Scheme#:~:text=The%20sample%20results%20show%20PFAS,supply%20is%20safe%20fo r%20use

WHO (2022). PFOS and PFOA in Drinking-water. Background document for development of WHO Guidelines for Drinking-water Quality. 29 September 2022. Version for public review. WHO/SDE/WSH/XXXXXX. World Health Organisation (WHO).

WSDH (2019a). Group A Public Water Supplies • Chapter 246-290 WAC. Draft Recommended State Action Levels for Per- and Polyfluoroalkyl Substances (PFAS) in Drinking Water: Approach, Methods and Supporting Information. November 2019. Washington State Department of Health (WSDH).

WSDH (2022b). Per- and Polyfluoroalkyl Substances Chemical Action Plan. Publication 21- 04-048. Revised September 2022. Washington State Department of Ecology. Washington State Department of Health (WSDH).

WSDH (2023a). 2023 EPA Proposal to Regulate PFAS in Drinking Water. 331-718. 3/15/2023. Washington State Department of Health (WSDH).

Xiao, F. (2022). An Overview of the Formation of PFOA and PFOS in Drinking-Water and Wastewater Treatment Processes. Journal of Environmental Engineering, 148(4), 01822001. https://doi.org/doi:10.1061/(ASCE)EE.1943-7870.0001986

Zhang Y., Beesoon S., Zhu L. and Martin J. W. (2013). Biomonitoring of perfluoroalkyl acids in human urine and estimates of biological half-life. Environmental Science & Technology 47(18): 10619-10627.

# **Appendix A List of Critical Studies Underpinning Guidance Value Derivation**

### **Evidence Evaluations for Australian Drinking Water Guidelines Chemical Fact Sheets – PFOS, PFHxS, PFOA, PFBS, and GenX Chemicals**

**PFOS, PFHxS, PFOA, PFBS, and GenX Chemicals Evaluation Report**

**National Health and Medical Research Council**

SLR Project No.: 640.V30693.20000

17 October 2024

The full list of critical studies underpinning each of the guidance values derived by various national and international jurisdictions (for which data extraction is provided in the accompanying Technical Report) is shown in **Table A-1** below, along with an indication of whether or not the critical study had been previously evaluated / considered by FSANZ (2017b, 2021). If they have been previously evaluated, the response to the question in the table 'Previously Evaluated / Considered by FSANZ?' would be 'Yes' and this is denoted with a tick (i.e.  $\vee$ ); conversely if the study(ies) have not been previously evaluated by FSANZ (2017b, 2021), the response to the question in the table would be 'No' and this is denoted with a cross (i.e.  $(x')$ ). Note the quidance values which have been subjected to further critical evaluation are those marked with a cross in the FSANZ (2017b) column (i.e. '<sup>x</sup>') in the **Table A-1**, i.e. those not previously evaluated / considered by FSANZ (2017b, 2021). If studies are marked with a tick (i.e.  $\sqrt{'}$ ) in that column, these critical studies have not been subjected to further evaluation in this report.

#### **Table A-1: List of critical studies underpinning each of the guidance values for the five PFAS covered in this review and indication of whether or not the critical study had been previously evaluated / considered by FSANZ (2017b, 2021)**







 $\checkmark$  = This study was previously evaluated / considered by FSANZ (2017b) or FSANZ (2021).  $\checkmark$  = This study has not been previously evaluated / considered by FSANZ (2017b) or FSANZ (2021).

Grey shading indicates the guidance value is based on an underpinning critical study which has not been previously evaluated / considered by FSANZ (2017b), and therefore has been further considered in this Evaluation Report (see also addendum to Research Protocol in **Section 3.4** of Technical Report).

#### **References for Appendix A**

Abraham K, Mielke H, Fromme H, Volkel W, Menzel J, Peiser M, Zepp F, Willich SN and Weikert C, 2020. Internal exposure to perfluoroalkyl substances (PFASs) and biological marker in 101 healthy 1-year-old children: associations between levels of perfluorooctanoic acid (PFOA) and vaccine response. Archives of Toxicology, 94, 2131–2147.

ATSDR (2021a). Toxicological Profile for Perfluoroalkyls. Released May 2021. Agency for Toxic Substances and Disease Registry (ATSDR).

BfR (2019a). New health-based guidance values for the industrial chemicals PFOS and PFOA BfR opinion No 032/2019 of 21 August 2019. German Federal Institute for Risk Assessment. Bundesinstitut für Risikobewertung (BFR).

Butenhoff J, Costa, G, Elcombe C, Farrar D, Hansen K, Iwai H, Jung R, Kennedy Jr. G, Lieder P, Olsen G and Thomford P (2002) Toxicity of Ammonium Perfluorooctanoate in Male cynomolgus Monkeys after Oral Dosing for 6 Months. Toxicol Sci 69: 244–257.

Butenhoff JL, Chang SC, Ehresman DJ, et al. (2009a). Evaluation of potential reproductive and developmental toxicity of potassium perfluorohexanesulfonate in Sprague Dawley rats. Reprod Toxicol 27:331-341. (as quoted in ATSDR 2021a).

Butenhoff JL, Chang S-C, Olsen GW and Thomford PJ (2012) Chronic dietary toxicity and carcinogenicity study with potassium perflurooctanesulfonate in Sprague Dawley rats. Toxicology 293: 1-15.

Budtz-Jørgensen, E., and P. Grandjean. 2018. Application of benchmark analysis for mixed contaminant exposures: mutual adjustment of perfluoroalkylate substances associated with immunotoxicity. PLoS One 13(10):e0205388.


Dong GH, Zhang YH, Zheng L, Liu W, Jin YH, He QC. (2009). Chronic effects of perfluorooctanesulfonate exposure on immunotoxicity in adult male C57BL/6 mice. Arch Toxicol. 83(9):805-815.

Dong, G., MM Liu, D Wang, L Zheng, ZF Liang, YH Jin, (2011). Sub-chronic effect of perfluorooctanesulfonate (PFOS) on the balance of type 1 and type 2 cytokine in adult C57BL6 mice. Archives of Toxicology 85: 1235-1244.

Dong GH, Zhang YH, Zheng L, Liang ZF, Jin YH, He QC. (2012a). Subchronic effects of perfluorooctanesulfonate exposure on inflammation in adult male C57BL/6 mice. Environ Toxicol. 27:285-296.

DuPont (2010). Oral (gavage) reproduction/developmental toxicity study in mice (OECD TG 421; modified according to the Consent Order) DuPont-18405-1037. Unpublished. As cited in MPART 2019a.

EFSA (2020). Risk to human health related to the presence of perfluoroalkyl substances in food. Adopted: 9 July 2020. European Food Safety Authority (EFSA).

Eriksen KT, Raaschou-Nielsen O, McLaughlin JK, Lipworth L, Tjønneland A, Overvad K, Sørensen M (2013): Association between plasma PFOA and PFOS levels and total cholesterol in a middle-aged Danish population. PLoS One. 2013;8(2):e56969 (as quoted in BfR 2019a).

Feng, X; Cao, X; Zhao, S; Wang, X; Hua, X; Chen, L; Chen, L. (2017). Exposure of pregnant mice to perfluorobutanesulfonate causes hypothyroxinemia and developmental abnormalities in female offspring. Toxicol Sci 155: 409-419.

FSANZ (2017b). Hazard assessment report – Perfluorooctane Sulfonate (PFOS), Perfluorooctanoic Acid (PFOA), Perfluorohexane Sulfonate (PFHxS). Food Standards Australia New Zealand (FSANZ).

Gallo, V., Leonardi, G., Genser, B., Lopez-Espinosa, M.J., Frisbee, S.J., Karlsson, L., Ducatman, A.M., Fletcher, T. (2012). Serum perfluorooctanoate (PFOA) and perfluorooctane sulfonate (PFOS) concentrations and liver function biomarkers in a population with elevated PFOA exposure. Environ Health Perspect 120(5): 655-660.

Grandjean, P., E.W. Andersen, E. Budtz-Jørgensen, F. Nielsen, K. Mølbak, P. Weihe, and C. Heilmann. 2012. Serum vaccine antibody concentrations in children exposed to perfluorinated compounds. JAMA 307:391–397 (as quoted in USEPA 2021d)

HC (2018a). Guidelines for Canadian Drinking Water Quality Guideline Technical Document Perfluorooctane Sulfonate (PFOS). December 2018. Health Canada (HC). Government of Canada.

Lau C, Thibodeaux JR, Hanson RG, Rogers JM, Grey BE, Stanton ME, Butenhoff JL and Stevenson LA (2003) Exposure to perfluorooctane sulfonate during pregnancy in rat and mouse. II: Postnatal evaluation. Toxicol Sci 74: 382–392.

Lau C, Thibodeaux JR, Hanson RG, Narotsky MG, Rogers JM, Lindstrom AB and Strynar MJ (2006) Effects of Perfluorooctanoic Acid Exposure during Pregnancy in the Mouse. Toxicol Sci 90 510–518.

Li K., Gao P., Xiang P., Zhang X., Cui X. and Ma L. Q. (2017). Molecular mechanisms of PFOA-induced toxicity in animals and humans: Implications for health risks. Environment International 99: 43-54.

Luebker, D.J., Case, M.T., York, R.G., Moore, J.A., Hansen, K.J., Butenhoff, J.L. (2005). Two-generation reproduction and cross-foster studies of perfluorooctanesulfonate (PFOS) in rats. Toxicol 215:126-148 (as quoted in ATSDR 2021a).



MDH (2020a). Toxicological Summary for: Perfluorooctane sulfonate. August 2020. Health-Based Guidance for Water. Health Risk Assessment Unit, Environmental Health Division. Minnesota Department of Health (MDH).

MDH (2020b). Toxicological Summary for: Perfluorohexane sulfonate. August 2020. Health-Based Guidance for Water. Health Risk Assessment Unit, Environmental Health Division. Minnesota Department of Health (MDH).

MDH (2022d). PFOA and Water. April 2022. Minnesota Department of Health (MDH).

MDH (2022e). PFBS and Drinking Water. March 2022. Minnesota Department of Health (MDH).

MDH (2022f). Toxicological Summary for: Perfluorooctanoate. March 2022. Health-Based Guidance for Water. Health Risk Assessment Unit, Environmental Health Division. Minnesota Department of Health (MDH)

MDH (2022g). Toxicological Summary for: Perfluorobutane sulfonate. March 2022. Health-Based Guidance for Water. Health Risk Assessment Unit, Environmental Health Division. Minnesota Department of Health (MDH).

MPART (2019a). Health-Based Drinking Water Value Recommendations for PFAS in Michigan. June 27, 2019. Michigan Science Advisory Workgroup. Michigan's PFAS Action Response Team (MPART).

Nelson JW, Hatch EE, Webster TF (2010): Exposure to polyfluoroalkyl chemicals and cholesterol, body weight, and insulin resistance in the general U.S. population. Environ Health Perspect 118(2):197-202 (as quoted in BfR 2019a).

NJDEP (2019a). Technical Support Document: Interim Specific Ground Water Criterion for Perfluorooctanoic Acid (PFOA, C8) (CAS #: 335-67-1; Chemical Structure: CF3(CF2)6COOH)\*. March 6, 2019. New Jersey Department of Environmental Protection (NJDEP).

NJDEP (2019b). Technical Support Document: Interim Specific Ground Water Criterion for Perfluorooctane Sulfonate (PFOS) (CAS #: 1763-23-1; Chemical Formula: C8HF17O3S). March 6, 2019. New Jersey Department of Environmental Protection (NJDEP).

NJDEP (2023a). Interim Specific Ground Water Quality Criterion (ISGWQC) for hexafluoropropylene oxide dimer acid (HFPO-DA) and its ammonium salt (GenX). May 24, 2023. Department of Environmental Protection. State of New Jersey (NJDEP).

NTP (2018). TOX-96: Toxicity Report Tables and Curves for Short-term Studies: Perfluorinated Compounds: Sulfonates and personal communication between MDH and NTP project manager Dr. Chad Blystone (as cited in the HRA Toxicology Review Worksheet for PFHxS, last revised 3/8/2019). National Toxicology Program (NTP).

NTP (2019). NTP Technical Report on the Toxicity Studies of Perfluoroalkyl Sulfonates (Perfluorobutane Sulfonic Acid, Perfluorohexane Sulfonate Potassium Salt, and Perfluorooctane Sulfonic Acid) Administered by Gavage to Sprague Dawley Rats P.H. Service, Editor. 2019, National Toxicology Program (NTP), U.S. Department of Health and Human Services: Research Triangle Park, NC.

NTP (2022). NTP technical report on the toxicity studies of perfluoroalkyl sulfonates (perfluorobutane sulfonic acid, perfluorohexane sulfonate potassium salt, and perfluorooctane sulfonic acid) administered by gavage to Sprague Dawley (Hsd:Sprague Dawley SD) rats (revised). Research Triangle Park, NC: National Toxicology Program. Toxicity Report 96.

OEHHA (2019a). Notification Level Recommendations. Perfluorooctanoic Acid and Perfluorooctane Sulfonate in Drinking Water. August 2019. Pesticide and Environmental



Toxicology Branch. Office of Environmental Health Hazard Assessment (OEHHA). California Environmental Protection Agency.

OEHHA (2021d). Notification Level Recommendation. Perfluorobutane Sulfonic Acid in Drinking Water. January 2021. Pesticide and Environmental Toxicology Branch. Office of Environmental Health Hazard Assessment (OEHHA). California Environmental Protection Agency.

OEHHA (2022a). Notification Level Recommendation. Perfluorohexane Sulfonic Acid in Drinking Water. March 2022. Pesticide and Environmental Toxicology Branch. Office of Environmental Health Hazard Assessment (OEHHA). California Environmental Protection Agency.

OEHHA (2023a). Public Health Goals. Second Public Review Draft. Perfluorooctanoic Acid and Perfluorooctane Sulfonic Acid in Drinking Water. July 2023. Pesticide and Environmental Toxicology Branch. Office of Environmental Health Hazard Assessment (OEHHA). California Environmental Protection Agency.

Onishchenko N, Fischer C, Wan Ibrahim WN, Negri S, Spulber S, Cottica D, Ceccatelli S. 2011. Prenatal exposure to PFOS or PFOA alters motor function in mice in a sex-related manner. Neurotox. Res. 19(3):452-61.

Perkins RG, Butenhoff JL, Kennedy Jr. GL, and Palazzolo MJ (2004) 13-Week Dietary Toxicity Study of Ammonium Perfluorooctanoate (APFO) in Male Rats. Drug Chem Toxicol 27 361–378.

RIVM (2021a). Revised Risk Assessment of GenX And PFOA in Food. Part 1: Toxicity of GenX and PFOA and intake through contaminated Dairy products, eggs and fish. 01-09- 2021 (final version). Rijksinstituut voor Volksgezondheid en Milieu (RIVM).

Seacat AM, Thomford PJ, Hansen KJ, Olsen GW, Case MT and Butenhoff JL (2002) Subchronic toxicity studies on perfluorooctanesulfonate potassium salt in cynomolgus monkeys. Toxicol Sci 68: 249-264.

Steenland K, Tinker S, Frisbee S, Ducatman A, Vaccarino V (2009): Association of perfluorooctanoic acid and perfluorooctane sulfonate with serum lipids among adults living near a chemical plant. Am J Epidemiol 170(10):1268-78 (as quoted in BfR 2019a).

Thibodeaux JR, Hanson RG, Rogers JM, Grey BE, Barbee BD, Richards JH, Butenhoff JL, Stevenson LA and Lau C (2003) Exposure to perfluorooctane sulfonate during pregnancy in rat and mouse. I: Maternal and prenatal evaluations. Toxicol Sci 74: 369-381.

Thomford PJ (2002) 104-week dietary chronic toxicity and carcinogenicity study with perfluorooctane sulfonic acid potassium salt (PFOS; T-6295) in rats. Final Report. Volumes I-IX. Covance study no. 6329-183. 3M Company, St Paul, MN.

USEPA (2021a). External Peer Review Draft. Proposed Approaches to the Derivation of a Draft Maximum Contaminant Level Goal for Perfluorooctanoic Acid (PFOA) (CASRN 335-67- 1) in Drinking Water. EPA Document No. 822D21001. December 2021. DRAFT. DO NOT CITE OR QUOTE. United States Environmental Protection Agency (USEPA).

USEPA (2021b). External Peer Review Draft. Proposed Approaches to the Derivation of a Draft Maximum Contaminant Level Goal for Perfluorooctane Sulfonic Acid (PFOS) (CASRN 1763-23-1) in Drinking Water. EPA Document No. 822D21002. December 2021. DRAFT. DO NOT CITE OR QUOTE. United States Environmental Protection Agency (USEPA).

USEPA (2021c). Human Health Toxicity Values for Perfluorobutane Sulfonic Acid (CASRN 375-73-5) and Related Compound Potassium Perfluorobutane Sulfonate (CASRN 29420-49- 3). EPA/600/R-20/345F. April 2021. United States Environmental Protection Agency (USEPA).

USEPA (2021e). Human Health Toxicity Values for Hexafluoropropylene Oxide (HFPO) Dimer Acid and Its Ammonium Salt (CASRN 13252-13-6 and CASRN 62037-80-3). Also Known as "GenX Chemicals". EPA-Final. EPA Document Number: 822R-21-010. October 2021. United States Environmental Protection Agency (USEPA).

USEPA (2022c). Technical Fact Sheet: Drinking Water Health Advisories for Four PFAS (PFOA, PFOS, GenX chemicals, and PFBS). EPA Document No. EPA 822-F-22-002. June 2022. United States Environmental Protection Agency (USEPA).

USEPA (2022d). Interim Drinking Water Health Advisory: Perfluorooctanoic Acid (PFOA) CASRN 335-67-1. EPA Publication # EPA/822/R-22/003. June 2022. United States Environmental Protection Agency (USEPA).

USEPA (2022e). Interim Drinking Water Health Advisory: Perfluorooctane Sulfonic Acid (PFOS) CASRN 1763-23-1. EPA Publication # EPA/822/R-22/004. June 2022. United States Environmental Protection Agency (USEPA).

USEPA (2022k). Drinking Water Health Advisory: Perfluorobutane Sulfonic Acid (CASRN 375-73-5) and Related Compound Potassium Perfluorobutane Sulfonate (CASRN 29420-49- 3). EPA/822/R-22/006. June 2022. United States Environmental Protection Agency (USEPA).

USEPA (2022j). Drinking Water Health Advisory: Hexafluoropropylene Oxide (HFPO) Dimer Acid (CASRN 13252-13-6) and HFPO Dimer Acid Ammonium Salt (CASRN 62037-80-3), Also Known as "GenX Chemicals". EPA-Final. EPA Document Number: EPA/822/R-22/005. June 2022. United States Environmental Protection Agency (USEPA).

USEPA (2023). IRIS Toxicological Review of Perfluorohexanesulfonic Acid (PFHxS, CASRN 335-46-4) and Related Salts. EPA Publication # EPA/635/R-23/148a. External Review Draft. July 2023. United States Environmental Protection Agency (USEPA).

WSDH (2019a). Group A Public Water Supplies • Chapter 246-290 WAC. Draft Recommended State Action Levels for Per- and Polyfluoroalkyl Substances (PFAS) in Drinking Water: Approach, Methods and Supporting Information. November 2019. Washington State Department of Health (WSDH).

WSDH (2022b). Per- and Polyfluoroalkyl Substances Chemical Action Plan. Publication 21- 04-048. Revised September 2022.

WSDH (2023a). 2023 EPA Proposal to Regulate PFAS in Drinking Water. 331-718. 3/15/2023. Washington State Department of Health (WSDH).



Making Sustainability Happen